

CHIEF EDITOR CO-CHIEF EDITORS

ASSOCIATE EDITORS

EXECUTIVE EDITORS

FRANCISC SCHNEIDER **IOANA SISKA** CARMEN TATU MIHAI NECHIFOR SORIN RIGA FLORINA BOJIN GABRIELATANASIE DACIANA NISTOR CALIN MUNTEAN

EDITORIAL BOARD

ARDELEAN AUREL **BADIU GHEORGHE** BĂDĂRĂU ANCA **BENEDEK GYORGY BENGA GHEORGHE BUNU CARMEN** COJOCARU MANOLE CUPARENCU BARBU CONSTANTIN NICOLAE HAULICĂ ION IANCAU MARIA MIHALAŞ GEORGETA MUNTEAN DANINA MUREŞAN ADRIANA NESTIANU VALERIU **OPREA TUDOR**

(Arad) (Constanța) (București) (Szeged) (Cluj) (Timişoara) (București) (Oradea) (Bucuresti) (laşi) (Craiova) (Timişoara) (Timişoara) (Cluj) (Craiova) (New Mexico)

PĂUNESCU VIRGIL (Timişoara) (Timişoara) PETROIU ANA POPESCU LAURENJIU (București) RÂCZ OLIVER (Kosice) **RIGA DAN** (București) SABĂU MARIUS (Tg. Mureş) SIMIONESCU MAIA (București) SIMON ZENO (Timişoara) SAULEA I. AUREL (Chisinău) SWYNGHEDAUW BERNARD (Paris) TANGUAY M. ROBERT (Canada) TATU FABIAN ROMULUS (Timişoara) VLAD AURELIAN (Timişoara) VOICU VICTOR (București) ZĂGREAN LEON (București)

ACCREDITED BY CNCSIS - B+CATEGORY CODE 240

http://journals.indexcopemicus.com/karta.php?action=masterlist&id=4929 http://www.ebscohost.com/titleLists/a9h-journals.pdf

Publication data: Fiziologia (Physiology) is issued quarterly	1. FOR SUBSCRIPTION ADDRESS
Subscription rates: Subscriptions run a full calendar year. Prices	HVB Bank TIMISOARA
are give per volume, surface postage induded.	RO 21 BACX 000000218508250
Personal subscription: Romania - 100 RON, Outside	
Romania - 35\$ (must be in the name of, billed to, and paid by an	TIMIŞOARA-ROMANIA
individual. Order must be marked "personal subscription")	PENTRU REVISTA
Instituțional subscription: 50\$ (regular rate)	"FIZIOLOGIA-PHYSIOLOGY"
Single issues and back volumes: Information on availability	
and prices can be obtained through the Publisher.	2.CORRESPONDENCE SHOULD BE ADDRESSED TO THE CHIEF EDITOR
Change of address: Both old and new address should be stated	ADDRESSED TO THE CHIEF EDITOR
and send to the subscription source.	PROF. DR. FRANCISC SCHNEIDER
Bibliographic indices: We hope this journal will be regularly listed	PO BOX 135
in bibliographic services, induding "Current Contents"	300024 - TIMIŞOARA - ROMANIA
Book Reviews: Books are accepted for review by special	e-mail: carmen.tatu@umft.ro
agreement.	
Advertising: Correspondence and rate requests should be	Editura EUROSTAMPA
addressed to the Publisher.	www.eurostampa.ro Bd. Revoluției din 1989 nr. 26, Timișoara
	Tel/fax: 0256-204816

Timisoara 6 ISSN 1223-2076

Instructions to Authors

Submission: Only original papers in English are corisidered and should be sentto:

Prof. dr. Francisc Schneider Chief Editor of "Fiziologia" PO Box 135 300024, TIMIŞOARA, ROMANIA TeUFax: 40-256/490507

Manuscripts should be submitted in triplicate sets of illustrations (of which one is an original), typewritten doublespaced on one side of the paper, with a wide margin.

Conditions: All manuscripts are subject to editorial review. Manuscripts are received with the explicit understanding that they are not under simultaneous consideration by any other publication. Submission of an artide for publication implies the transfer of the Copyright from the authorto the publisher upon acceptance. Accepted papers become the permanent property of "Fiziologia" (Physiology) and may not be reproduced by any means, in-whole or in part, without the written consent of the publisher. It is the author's responsibility to obtain permission to reproduce illustrations, tables, etc. from other publications.

Arrangement:

Title page: The first of each paper should indicate the title (main title underlined), the authors' names, and the institute where the work was conducted. A short title for use as running head is also required.

Keywords: for indexing purposes, a list of 3-10 keywords in English and Romanian is essential.

Abstract: Each paper needs abstract and title in Romanian and English language, fonts size 9, Arial Narrow.

Bady text: fonts size 10, Arial Narrow.

Small type: Paragraphs which can or must be set in smaller type (case histories, test methods, etc.) should be indicated with a "p" (petit) in the margin on the left-hand side.

Footnotes: Avoid footnotes. When essential, they are numbered consecutively and typed at the foot of the appropriate page, fonts size 8, Arial Narrow.

Tables and illustrations: Tables (numbered in Roman numerals) and illustrations (numbered in Arabic numerals) should be prepared on separate sheets, fonts size 9, Arial Narrow. Tables require a heading, and ligures a legend, also prepared on a separate sheet. For the reproduction of illustrations, only good drawings and original photographs can be accepted; negatives or photocopies cannot be used. When possible, group several illustrations on one blockfor reproduction (max. size 140x188 mm) or provide crop marks. On the back of each illustration indicate its number, the author's name,

and artide title. Colour illustration are reproduced at the author's expense.

References: In the text identify references by Arabic figures, (in brackets), fonts size 9, Arial Narrow. Material submitted for publication but not yet accepted should be noted as "unpublished data" and not be induded in the reference list. The list of references should include only those publications which are cited in the text. The references should be numbered and arranged alphabetically by the authors' names. The surnames of the authors followed by initials should be given. There should be no punctuation signs other than a comma to separate the authors. When there are more than 3 authors, the names of the 3 only are used, followed by "et al" abbreviate journal names according to the Index Medicus system. (also see International Committee of Medical Journal Editors: Uniform Requirements for manuscripts submitted to biomedical journals. Ann Intern Med 1982; 96: 766-771).

Examples:

(a) Papers published in perfodicals: Kauffman HF, van der Heide S, Beaumont F, et al: Class-apecific antibody determination against Aspergillus fumigatus by mean of the enzyme-linked immunosorbent assay. III. Comparative study: IgG, IgA, IgM, ELISA titers, precipitating antibodies and IGE biding after fractionation of the antigen. Int Arch Allergy Appl Immunol 1986; 80:300 - 306.

(b) Monographs; Matthews DE, Farewell VT: Using and Understanding Medical Statistics. Basel, Karger, 1985.

(c) Edited books: Hardy WD Jr, Essex M:.FeLV-inducted feline acquired immune deficiency syndrome: A model for human AIDS; in Klein E(ed): Acquired Immunodeficiency Syndrome. Prag Allergy, Busel, Karger, 1986, vol 37,353 - 376.

Full address: The exact postai address complete with postai code of the senior author must be given; if correspondence is handled by someone else, indicate this accordingly. Add the E-mail address if possible.

Page charges: There is no page charge for papers of 4 or fewer printed pages (induding tables, illustrations and references).

Galley proofs: unless indicated otherwise, galley proofs are sent to the first-named authOr and should be returned with the least possible delay. Alternations made in galley proofs, other than the corrections of printer's errors, are charged to the author. No page proofs are supplied.

Reprints: Order forms and a price list are sent with the galley proofs. Orders submitted after the issue is printed are subject to considerably higher prices. Allow five weeks from date of publication for delivery of reprints.



CONTENTS

1. Effect of Magnegita on Selected Blood Parameters Minca DM, Brad S, Tatu C, Cobzariu Fl	4
2. Analysis of Sterolic Compounds from Vegetal Extracts of Ribes Nigrum, Rosa Canina, Betula Pubescens, Carpinus Betulus, Viburnum Lantana and Propolis Tincture Orodan M, Vlase L, Istudor V	10
3. Antibacterial Effects of Some Plant Extracts on Staphylococccus Aureus Strains Simon LM, Pepelea L, Junie LM	16
4. Acute Oral and Dermal Effects of Spirulina in Mice Andrica FM, Dehelean C, Serban MC, Pânzaru I, Coricovac D, Drăgan S	22
5. The Role of Spirulina Platensis in the Control of Type 2 Diabetes Mellitus Serban MC, Stoichescu-Hogea G, Gurban C, Petcu F, Jeyakumar D, Andrica F, Munteanu C, Dragan S	27
6. Effects of Resveratrol and Coenzyme Q10 Supplementation in Metabolic Syndrome Ardelean F, Stoichescu-Hogea G, Gurban C, Şerban MC, Petcu F, Antal DS, Drăgan S	32
7. Novel Insights into the Role of Methylene Blue in Mitochondrial Protection Duicu OM, Sturza A, Scurtu-Mytiko I, Privistirescu A, Dănilă M, Noveanu L, Muntean DM	38
CUPRINS	
1. Efectul Magnegita asupra unor parametri sanguine Minca DM, Brad S, Tatu C, Cobzariu Fl	4
2. Analiza compusilor sterolici din extractele vegetale de Ribes Nigrum, Rosa Canina, Betula Pubescens, Carpinus Betulus, Viburnum Lantana și tinctura de propolis Orodan M, Vlase L, Istudor V	10
3. Efectele antibacteriene ale unor extracte din plante asupra tulpinilor de <i>Staphylococccus Aureus</i> Simon LM, Pepelea L, Junie LM	16
4. Efectele acute orale și cutanate ale spirulinei pe model animal de șoarece Andrica FM, Dehelean C, Serban MC, Pânzaru I, Coricovac D, Drăgan S	22
5. Rolul Spirulina Platensis în controlul diabetului zaharat tip 2 Serban MC, Stoichescu-Hogea G, Gurban C, Petcu F, Jeyakumar D, Andrica F, Munteanu C, Dragan S	27
6. Efectele suplimentelor alimentare conținând resveratrol și coenzima Q10 la pacienții cu sindrom metabolic Ardelean F, Stoichescu-Hogea G, Gurban C, Şerban MC, Petcu F, Antal DS, Drăgan S	32

EFFECT OF MAGNEGITA ON SELECTED BLOOD PARAMETERS

DELIA MARIA MINCA¹, SILVIU BRAD², CARMEN TATU², FLORIN IOSIF COBZARIU²

¹Pius Branzeu County Clinical Emergency Hospital Timisoara ²Victor Babes University of Medicine and Pharmacy Timisoara

ABSTRACT

Magnetic resonance imaging (MRI) is a noninvasive diagnostic test widely used in clinical practice. Signal resolution and sensitivity can be further enhanced by adding contrast agents in order to make abnormalities or disease processes more visible. Gadopentetate dimeglumine was the first approved gadolinium-based contrast agent (GBCA) and showed good diagnostic value and high levels of safety and tolerance. However, if the free gadolinium (Gd) ion is released *in vivo*, cardiovascular and neurologic toxicity may occur. The plasma half-life of the Gd chelates is dependent upon the volume of distribution and the glomerular filtration rate of the agent, and it is higher in patients with impaired renal function. This is why both FDA and EMEA recommended caution when using GBCA-enhanced MRI in patients with moderate renal impairment and banned their use in those with severe renal impairment. The aim of this study was to assess the effect of Magnegita on certain laboratory parameters in patients with normal renal function undergoing cerebral and abdominal MRI scans. Twenty-one patients were included from whom we collected three blood samples (prior to MRI scan, at 30 minutes and 24 hours after). We have assessed 27 parameters by the means of hematology, biochemistry and ionogram tests. Our results showed minor and transient changes in some of the parameters, but most of them remained unchanged throughout the study. Further studies are needed to determine the effect of gadopentetate dimeglumine on the same laboratory parameters in patients with impaired renal function. **Key words**: Magnegita, gadopentetate dimeglumine, laboratory parameters

INTRODUCTION

Magnetic resonance imaging (MRI) is a noninvasive diagnostic test widely used in clinical practice as it allows for the simultaneous examination of the anatomy and physiological parameters of tissues and organs by measuring the radio-frequency signals emitted by water protons in living tissues, without using harmful radiation [1,2]. In order to make abnormalities or disease processes more visible, the signal resolution and sensitivity need to be enhanced and this is achieved by adding contrast agents (CAs); therefore approximately 35% of MRI scans are currently performed with the use of CAs [3, 4]. Gadolinium (Gd), a rare-earth metal of the lanthanide series, is the most widely used metal for the production of CAs, being administered in the form of chelate complexes in order to prevent the release of toxic metal ion [5] The free gadolinium ion has cardiovascular and neurologic toxicity, being deposited in liver, bone and lymph nodes [6].

Gadopentetate dimeglumine (Gd-DTPA) was the first approved gadolinium-based contrast agent (GBCA) [7,8]. Early studies [9-11] evaluating the safety and efficacy of Gd-DTPA have concluded it has higher diagnostic value compared to the non contrast-enhanced MRI and exhibits high levels of safety and tolerance. In a multicenter double-blind randomized clinical trial [9], Gd-DTPA improved diagnostic ability in 65% of patients and led to changes in presumptive diagnosis, along with visualization of more lesions in some of those patients. Adverse reactions in some patients were reported, such as asymptomatic rise in serum iron and bilirubin levels, hyper- or hypotension, weakness, conjunctivitis, local burning at injection site, headache, nausea and vomiting, all of which were minor and transient. These adverse reactions were comparable to those related to administration of iodinated non-ionic roentgen contrast media [12].

However, the plasma half-life of the Gd chelates is dependent upon the volume of distribution and the glomerular filtration rate of the agent, and it is higher in patients with impaired renal function [13]. Contrast-induced nephropathy, defined as increase in creatinine concentration [14] or decrease in glomerular filtration rate [15] has been associated with GBCAs, particularly at high doses and in patients with renal impairment. Moreover, nephrogenic systemic fibrosis (NSF), a life-threatening fibroproliferative

Received 10th of May 2015. Accepted 5th of June 2015. **Address for correspondence**: Delia Minca, MD, PhD student, Pius Branzeu County Clinical Emergency Hospital Timisoara, Iosif Bulbuca No. 10 Street, RO-300736, Timisoara, Romania; phone: +40356433121; fax: +40256486956; e-mail: delia.minca@yahoo.com

disease seen in people with severe renal impairment, was first observed in 1997 [16] and it has been associated with the use of GBCAs. NSF is thought to be caused by the deposition of free gadolinium in tissues of patients with severe renal dysfunction and the subsequent inability of kidneys to eliminate it because of poor water solubility of the gadolinium [17]. Consequently, in 2007 the USA Food and Drug Administration requested manufacturers to add a warning box to the labels of all GBCAs, stating the increased risk of NSF in patients with acute or chronic severe renal insufficiency [18], while the European Medicines Agency (EMEA) has released in 2009 a set of recommendations [19], classifying the active substances in the GBCAs into three categories of risk: high-, medium-, and low-risk. EMEA's Committee for Medicinal Products for Human Use has recommended contraindications in patients with severe kidney problems, in those scheduled for or who had recently received a liver transplant and in newborn babies up to four weeks of age for high-risk gadolinium-containing contrast agents, including Magnegita. Additionally, patients with unknown kidney problems should always be screened for kidney problems using laboratory tests before the use of a GBCA [19]. Finally, a wide variety of changes in physiology may occur following the administration of gadolinium salts and chelates [20, 21]. However, most of the studies have focused on the effects of GBCAs on kidneys, while just a few have specifically addressed the physiological changes triggered by these agents.

The aim of this study was to assess the effect of Magnegita on certain laboratory parameters in patients with normal renal function undergoing cerebral and abdominal MRI scans.

MATERIALS AND METHODS

The study group was composed of 21 patients who underwent cerebral and abdominal MRI scans in the Radiologica Plus Medical Imagistics Centre Timisoara in the period January - March 2015. No patient has had kidney or liver impairment.

Inclusion criteria: patients aged 20-60, both genders; not pregnant or breastfeeding, in case of women; had not had an organ transplant; had not received any medications within 48 hours of participation; had not received any contrast agent within 7 days; willing to sign Informed Consent Form; willing to provide three blood samples.

Exclusion criteria: patients with impaired renal function; patients who have had a contrast-enhanced MRI or CT seven days prior to inclusion in the study; under chemoor radiation therapy; highly elevated or decreased laboratory parameters.

Most of the patients (18; 7 women, 11 men) were under

40 years of age, while the other three (2 women, 1 man) were aged over 40 (Figure 1). The number of men included was slightly higher (Figure 2).

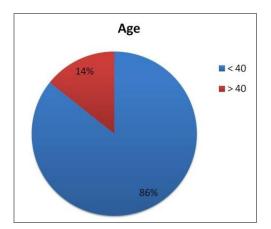


Fig. 1. Distribution of patients by age group

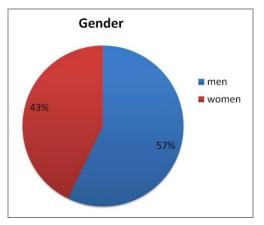


Fig. 2. Distribution of patients by gender

All the patients in this study received 0.1 ml/kg body weight gadopentetate dimeglumine (Magnegita ® 500 micromol/ml, Biokanol Pharma GmbH) by intravenous injection once and the exposure to the contrast agent was not repeated in any of the patients. As the use of GBCAs has been linked to the risk of developing nephrogenic systemic fibrosis, all the patients are screened for renal dysfunction prior to be administered Magnegita. None of the patients included in our study suffered from moderate (GFR 30–59 ml/min/1.73 m²) or severe (GFR < 30 ml/min/1.73m²) renal impairment.

After the patients have signed the Informed Consent Form to be included in this study, two blood samples of 3 ml (for complete blood count) and 6 ml (for biochemistry tests), respectively, were collected from each patient for three times, namely prior to MRI, then 30 minutes and 24 hours after the MRI scan. We have analysed the same 27 parameters (see Table I) by the mean of complete blood count, biochemistry and ionogram tests. We have used reagents from Diagon for hematology tests, DiaSys Diagnostic Systems for biochemistry tests and Medica for ionogram tests. All reagents were used according to the manufacturer's instructions.

Type of laboratory test	Method/Equipment	Parameter			
		white blood cell count (WBC)			
		neutrophils (absolute count; percentage)			
		lymphocytes (absolute count; percentage)			
		monocytes (absolute count; percentage)			
		eosinophils (absolute count; percentage)			
		basophils (absolute count; percentage)			
		red blood cell count (RBC)			
Complete blood	mean corpuscular h mean corpuscular h concentration (MCH	hemoglobin (HGB)			
count		hematocrit (HCT)			
		mean corpuscular volume (MCV)			
		mean corpuscular hemoglobin (MCH)			
		mean corpuscular hemoglobin			
		concentration (MCHC)			
		red cell distribution width (RDW)			
		platelets (PLT)			
		concentration (MCHC) red cell distribution width (RDW)			
		serum urea			
		serum creatinine			
Biochemistry	Spectrophotometry /	glycemia			
Diochemistry	HITACHI 917	alanine aminotransferases (ALT)			
		aspartate aminotransferases (AST)			
		total serum proteins			
		serum calcium			
	Ion selective electrode	serum magnesium			
lonogram	technology / EasyLyte	serum sodium			
	Plus	serum potassium			
		serum chloride			

RESULTS

At baseline, the WBC level was normal in 20 patients and lower-than-normal in one; no changes were seen in the WBC level of the latter patient following the administration of the contrast agent at the subsequent blood tests. Thirty minutes after the injection of Magnegita, the WBC level slightly increased in one patient and then returned to normal after 24 hours. Additionally, a slightly decreased level was recorded in one patient at 24 hours, although this patient has had normal values both at baseline and 30 minutes post-injection. As mentioned above, the changes were minor, with no clinical significance.

The white blood cell differential has yielded various results. The absolute count of neutrophils, lymphocytes and monocytes has remained unchanged throughout the three measurements. However, the neutrophil percentage was above the normal range limit in three patients at baseline and it remained high after the contrast agent was injected; the only difference noted was in one patient with lower values at 30 minutes and 24 hours. The lymphocyte percentage was in the normal range in 16 patients at baseline (lower values - 3 patients; higher values -2 patients); 30 minutes post-injection, the lymphocyte percentage has reached the normal range in

one of the patients who had subnormal values prior to injection of Magnegita, but dropped again at 24 hours, when we also found one more patient with increased values. Distribution of patients with lower, normal or higher neutrophil and lymphocyte percentages across the three measurements is shown in Figure 3.

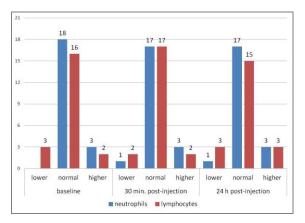


Fig. 3. Distribution of patients according to neutrophil and lymphocyte percentages

The absolute count of eosinophils revealed 11 patients had normal values at baseline, but only five after 30 minutes and 24 hours, respectively, while the other patients (10 at baseline, 16 at the other two measurements) had slightly decreased levels. Nevertheless, the eosinophil percentages exhibited values in the normal range for 17 patients (higher - 4 patients) at the baseline, for 19 patients (higher - 2 patients) 30 minutes after the administration of the GBCA, and for 18 patients (higher - 3 patients) 24 hours post-injection (Figure 4).

Both the absolute count and the percentage of basophils were in the normal range for 20 patients (higher - 1 patient) at the first and second measurements, while at the third measurement, values of 19 patients (higher - 2 patients) remained normal. The monocyte percentage has shown no variations, with one of the patients having decreased values throughout the study.

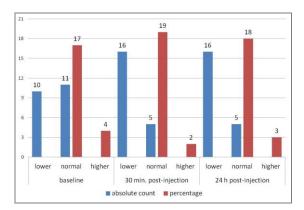


Fig. 4. Distribution of patients according to absolute count and percentage of eosinophils

Red blood cells count showed three patients had lower levels at baseline and remained unchanged at the other two measurements, while the other 18 patients had normal levels. After the injection of Magnegita, RBC dropped in another three patients at the 30 minutes post-injection measurement, but returned to normal after 24 hours. We have also found one patient with higher levels at the 24 h blood test (see Fig. 5). Lower levels of hemoglobin and hematocrit were also seen in four and five patients, respectively, at all three measurements. In three of the patients who had normal levels at baseline, HGB has decreased 30 minutes post-injection, but returned to normal after 24 hours. Similarly, the HCT level has decreased at the second measurement in two patients, but it was normal after 24 hours. The mean platelet volume was higher in three patients at baseline, but it decreased in one of them to a value within the normal range at the subsequent measurements. The platelet distribution width has slightly decreased in one patient 24 h after the injection of the contrast agent. No changes were recorded in mean corpuscular volume, mean corpuscular hemoglobin, mean corpuscular hemoglobin concentration, red cell distribution width, and platelets.

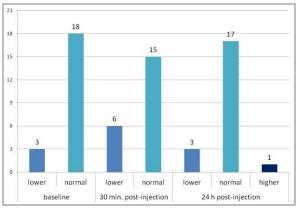


Fig. 5. Distribution of patients according to RBC levels

The biochemistry tests showed no changes in serum urea, serum creatinine, glycemia, and aspartate aminotransferases. Two patients had higher-than-normal levels of alanine aminotransferases at baseline, but only one at the following measurements. The total serum proteins were also higher at baseline in three patients, and then decreased in two of them at the subsequent tests.

The serum calcium level was higher in one patient throughout the study. At the second measurement, it decreased in another patient, while 24 hours later two patients had lower levels (Figure 6). The serum chloride was found to be higher than normal in 12 patients at baseline, and then in 13 and 14 patients at 30 minutes and 24 hours, respectively, after the injection of Magnegita (Figure 7). Serum magnesium, sodium and potassium remained unchanged during the study.

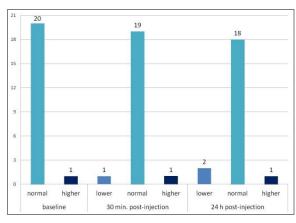


Fig. 6. Distribution of patients according to serum calcium levels

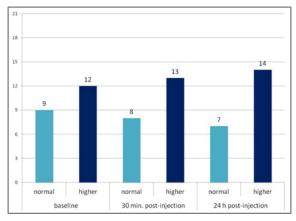


Fig. 7. Distribution of patients according to serum chloride levels

DISCUSSION

Gadolinium chelates are routinely used nowadays in magnetic resonance imaging, mostly for CNS pathology, followed by contrast-enhanced MR angiography, liver imaging, and breast evaluation for malignancy [22]. After intravenous injection, they rapidly distribute in the blood and the extracellular space [13, 23] and are normally eliminated unmetabolized via kidneys through glomerular filtration [13, 24], with half-lives of approximately 1.5 hours in people with normal renal function [5, 13, 23]. Unfortunately, toxic Gd(III) ions may be released in vivo by transmetallation with other metal ions, thus the kinetic stability of the gadolinium chelates is very important [3]. The ionic linear Gd chelates (such as gadopentetate dimeglumine) were shown to be more stable than the nonionic linear ones under normal and particularly under altered physiological conditions [25]. Although early studies have found gadopentetate dimeglumine is safe and well tolerated [9-11] in people with normal renal function, it has been later proved that GBCAs may trigger nephrogenic systemic fibrosis in patients with kidney disorders [6, 26, 27]. Therefore, for safety reasons, the benefits and risks of GBCA-enhanced MRI should be carefully balanced in patients with impaired renal function and also in patients with liver impairment.

A double-blind, randomized, crossover, placebo-controlled study designed to evaluate the safety and tolerance of gadopentetate dimeglumine 0.1 mmol/kg when administered to normal subjects at a four times higher bolus injection rate than that recommended by the producer found no clinically significant changes in laboratory parameters, even a few values outside the specified normal ranges were recorded [28].

Nevertheless, other data in the literature show gadolinium may interfere with various laboratory parameters, although changes are minor and transient. In a retrospective study, data regarding serum calcium, magnesium and creatinine levels were reviewed before and within 1 day after gadodiamide or gadopentetate dimeglumine administration [29]. A transient decrease in serum creatinine was seen in patients with renal impairment who received gadopentetate dimeglumine, but no changes were recorded in serum calcium and magnesium. Conversely, 36.7% of the patients with high-dose gadodiamide injection and renal insufficiency had spurious critical hypocalcemia and minor increases in serum magnesium. Moreover, other in vivo and in vitro studies have documented the interference with gadodiamide colorimetric of assays and gadoversetamide, resulting in spurious hypocalcemia, but gadopentetate dimeglumine has not been shown to cause this interference [20, 30].

The study by Proctor et al. [20] on the effects of gadoversetamide, gadodiamide, gadopentetate dimeglumine, and gadoteridol has shown that GBCAs can produce positive or negative analytic interference with multiple serum assays, including angiotensin-converting enzyme, calcium. iron. magnesium, total iron binding capacity, and zinc, with the first two producing the most interference. Gadopentetate dimeglumine showed interference with iron, while a manual colorimetric zinc assay showed interference with all the GBCAs studied. However, the other parameters measured (41 in total, including some of the parameters we have studied) showed no significant interference with any of the four GBCAs.

Our study results are mostly consistent with these reports. Furthermore, we have found minor and transient or no changes in the other parameters evaluated in this study.

CONCLUSION

Although the risk of developing nephrogenic systemic fibrosis or contrast-induced nephropathy is currently a concern in people with impaired renal function, the use of GBCAs in patients with normal renal function remains safe. Moreover, gadopentetate dimeglumine is among the safest GBCAs and it triggered no clinically significant changes in the laboratory parameters of the patients included in this study. Further studies are needed to determine the effect of gadopentetate dimeglumine on the same laboratory parameters in patients with impaired renal or hepatic function.

REFERENCES

1. Keevil SF. Magnetic resonance imaging in medicine. *Physics Education* 2001; 36(6): 476-485.

2. Strijkers GJ, Mulder M, Willem J, van Tilborg F, Geralda A and Nicolay K. MRI contrast agents: current status and future perspectives. *Anti-Cancer Agents in Medicinal Chemistry* 2007; 7(3): 291-305.

3. Hermann P, Kotek J, Kubicek V, Lukes I. Gadolinium(III) complexes as MRI contrast agents: ligand design and properties of the complexes. *Dalton Trans* 2008; 23: 3027-3047.

4. Major JL, Meade TJ. Bioresponsive, cell-penetrating, and multimeric MR contrast agents. *Acc Chem Res* 2009; 42: 893-903.

5. Werner EJ, Datta A, Jocher CJ, Raymond KN. High-relaxivity MRI contrast agents: Where coordination chemistry meets medical imaging. *Angew. Chem. Int. Ed.* 2008; 47: 8568-8580.

6. Penfield JG, Reilly RF. Nephrogenic systemic fibrosis and the use of gadolinium-based contrast agents. *Pediatr Nephrol.* 2007; 3(12): 654-668.

7. Frullano L, Caravan P. Strategies for the preparation of bifunctional gadolinium(III) chelators. *Curr. Org. Synth.* 2011; 8(4): 535–565

8. Zhou Z, Lu ZR. Gadolinium-based contrast agents for magnetic resonance cancer imaging. *Nanomed. Nanobiotechnol.* 2013; 5(1): 1-18.

9. Russell EJ, Schaible TF, Dillon W, *et al.* Multicenter double-blind placebo-controlled study of gadopentetate dimeglumine as an MR contrast agent: evaluation in patients with cerebral lesions. *AJNR* 1989; 10: 53-63.

10. Goldstein HA, Kashanian FK, Blumetti RF, Holyoak WL, Hugo FP, Blumenfield DM. Safety assessment of gadopentetate dimeglumine in U.S. clinical trials. *Radiology* 1990; 174: 17-23.

11. Carollo B, Runge VM, Price AC, Nelson KL, Wolf CR, Pacetti MI. The prospective evaluation of Gd-DTPA in 225 consecutive cranial cases: adverse reactions and diagnostic value. *Magn Reson Imaging* 1990; 8: 381-393.

12. Niendorf HP. Tolerance and safety of Gd-DTPA in 7000 patients: a review. *Diagnostic Imaging Internationa*11988; 4(S): 15-20.

13. Ersoy H, Rybicki FJ: Biochemical safety profiles of gadolinium-based extracellular contrast agents and nephrogenic systemic fibrosis. *J Magn Reson Imaging* 2007; 26(5): 1190–1197.

14. Briguori C, Colombo A, Airoldi F, *et al.* Gadolinium-based contrast agents and nephrotoxicity in patients undergoing coronary artery procedures. *Catheterization and Cardiovascular Interventions* 2006; 67(2): 175-180.

15. Erley CM, Bader BD, Berger ED, *et al.* Gadolinium-based contrast media compared with iodinated media for digital subtraction angiography in azotaemic patients. *Nephrology Dialysis Transplantation* 2004; 19(10): 2526-2531.

16. Tavernaraki A, Skoula A, Benakis S, and Exarhos D. Side Effects and Complications of Magnetic Resonance Contrast Media. *Hospital Chronicles* 2012; 7(4): 208-214.

17. Penfield JG and Reilly RF. What nephrologists need to know about gadolinium. *Nature Clinical Practice Nephrology* 2007; 3(12): 654-668.

 http://www.fda.gov/downloads/safety/medwatch/safetyinf ormation/safetyalertsforhumanmedicalproducts/ucm154532.pdf
 http://www.ema.europa.eu/docs/en_GB/document_library

/Press_release/2009/11/WC500015569.pdf

20. Proctor KAS, Rao LV and Roberts WL. Gadolinium magnetic resonance contrast agents produce analytic interference in multiple serum assays. *American Journal of Clinical Pathology* 2004; 121(2): 282-292.

21. Adding LC, Bannenberg GL, Gustafsson LE. Basic experimental studies and clinical aspects of gadolinium salts and chelates. *Cardiovascular drug reviews* 2001; 19(1):41-56.

22. Runge VM. Current Technological Advances in Magnetic Resonance with Critical Impact for Clinical Diagnosis and Therapy. *Investigative Radiology* 2013; 48(12): 869-877.

23. Aime S, Caravan P. Biodistribution of gadolinium-based contrast agents, including gadolinium deposition. *J Magn Reson Imaging.* 2009; 30: 1259–67.

24. Canga A, Kislikova M, Martinez-Galvez M et al. Renal function, nephrogenic systemic fibrosis and other adverse

reactions associated with gadolinium-based contrast media. *Nefrologia* 2014; 34(4): 428-38.

25. Frenzel T, Lengsfeld Ph, Schimer H, Hütter J, and Weinmann HJ. Stability of Gadolinium-Based Magnetic Resonance Imaging Contrast Agents in Human Serum at 37 [degrees] C. *Investigative Radiology* 2008; 43(12): 817-828.

26. Thomsen HS, Marckmann P, Logager VB. Nephrogenic systemic fibrosis (NSF): a late adverse reaction to some of the gadolinium based contrast agents. *Cancer Imaging* 2007; 7: 130-137.

27. Zou Z, Zhang H, Roditi GH, Leiner T, Kucharczyk W, and Prince MR. Nephrogenic systemic fibrosis: review of 370 biopsy-confirmed cases. *JACC: Cardiovascular Imaging* 2011; 4(11): 1206-1216.

28. Kashanian FH, Goldstein HA, Blumetti RF, Holyoak WF, Hugo FP, Dolker M. Rapid bolus injection of gadopentetate dimeglumine: absence of side effects in normal volunteers. *American Journal of Neuroradiology* 1990;11(5): 853-856.

29. Zhang HL, Ersoy H, Prince MR. Effects of gadopentetate dimeglumine and gadodiamide on serum calcium, magnesium, and creatinine measurements. *J Magn Reson Imaging* 2006;23(3):383-387.

30. Moore CD, Newman RC, Caridi JG. Spurious hypocalcemia after gadodiamide-enhanced magnetic resonance imaging: a case report and review of the literature. *Reviews in Urology* 2006; 8(3):165.

EFECTUL MAGNEGITA ASUPRA UNOR PARAMETRI SANGUINI

REZUMAT

Imagistica prin rezonantă magnetică (IRM) este o metodă non-invazivă de diagnostic larg utilizată în pracica clinică. Rezolutia si sensibilitatea semnalelor poate fi sporită prin adăugarea de agenti de contrast, cu scopul de a creste vizibilitatea anomaliilor sau a proceselor patologice. Gadopentetatul de dimeglumină a fost primul agent de contrast pe bază de gadoliniu (GBCA) aprobat si a dovedit o valoare diagnostic bună, precum si un nivel ridicat de sigurantă și tolerabilitate. Totuși, dacă ionul liber de gadoliniu (Gd) este eliberat in vivo, poate apărea toxicitatea cardio-vasculară și neurologică. Timpul de înjumătătire în plasmă a chelatilor de Gd depinde de volumul de distribuție și de rata de filtrare glomerulară a fiecărui agent de contrast și este mai mare la pacienții cu suferință renală. De aceea, atât FDA, cât și EMEA au recomandat prudență în utilizarea IRM cu GBCA la pacientii cu insuficientă renală moderată si au interzis utilizarea lor la cei cu insuficientă renală severă. Scopul acestui studiu a fost de a evalua efectul Magnegita asupra anumitor parametri de laborator la pacienți cu funcție renală normală care au fost evaluați prin IRM cerebral sau abdominal. În studiu au fost incluși 21 de pacienți, de la care s-au recoltat trei probe de sânge (înainte de IRM, la 30 de minute și la 24 de ore după). Am evaluat 27 de parametri prin intermediul testelor de hematologie, biochimie si ionogramă. Rezultatea noastre au arătat că unii parametri au suferite modificări minore si tranzitorii, majoritatea rămânând neschimbati. Sunt necesare alte studii pentru a determina efectul pe care gadopentetatul de dimeglumină îl are asupra parametrilor de laborator la pacientii cu disfuncție renală.

Cuvinte cheie: Magnegita, gadopentetat de dimeglumină, parametri de laborator

ANALYSIS OF STEROLIC COMPOUNDS FROM VEGETAL EXTRACTS OF *RIBES NIGRUM*, *ROSA CANINA*, *BETULA PUBESCENS*, *CARPINUS BETULUS*, *VIBURNUM LANTANA* AND PROPOLIS TINCTURE

MIHAELA ORODAN¹, LAURIAN VLASE², VIORICA ISTUDOR³

¹Clinical Emergency Hospital Arad and "Carol Davila" University of Medicine and Pharmacy Bucharest ²Deptartment of Pharmaceutical Technology and Biopharmaceutics, Faculty of Pharmacy, University of Medicine and Pharmacy "Iuliu Hatieganu" Cluj-Napoca

³"Carol Davila" University of Medicine and Pharmacy Bucharest

ABSTRACT

Gemmotherapic vegetal extracts in DH1 dilution of *Ribes nigrum*, *Rosa canina*, *Betula pubescens*, *Carpinus betulus*, *Viburnum lantana*, 30% propolis tincture were analyzed through high performance chromatography coupled with mass spectrometry (LC/MS) to analyze the sterols, specifically: betasitosterol, stigmasterol, campesterol, ergosterol, brassicasterol.

Gemmotherapic vegetal extracts in the DH1 dilution of *Rosa canina* (X1), *Betula pubescens* (X2), *Ribes nigrum* (X3), *Viburnum lantana* (X5), *Carpinus betulus* (X6) and propolis tincture 30% (X4) are frequently used in integrative medicine for their anti-inflammatory properties. Due to the fact that the anti-inflammatory effect of the vegetal extracts may also be induced by the sterols, we proposed to analyze these extracts (obtained from Plantextract, Cluj), through high performance chromatography coupled with mass spectrometry (LC/MS)[1,2,3]. The extracts analyzed were betasitosterol, stigmasterol, campesterol, ergosterol, brassicasterol [1,3]. The concentrations of sterols found through the method above mentioned (expressed in μ g/mL) were significant only for betasitosterol in: X1 = 15.859, X2 = 17.251, X3 = 21.598, X4 = 1.516; X5 = 2.033, X6 - unidentifiable.

Keywords: Gemmotherapic vegetal extracts, sterols and propolis, LC/MS/MS analysis

INTRODUCTION

Gemmotherapic vegetal extracts in the DH1 dilution of *Rosa canina* (X1), *Betula pubescens* (X2), *Ribes nigrum* (X3), *Viburnum lantana* (X5), *Carpinus betulus* (X6) and propolis tincture 30% (X4) are frequently used in integrative medicine for their anti-inflammatory and antiallergic properties. Due to the fact that we didn't find research regarding their chemical composition in specialty literature, in this study we analyzed the sterol compounds using high performance chromatography coupled with mass spectrometry (LC/MS), as they are known for anti-inflammatory effect.

MATERIALS AND METHODS

Devices: HPLC coupled with HP1100 Series mass spectrometer with binary pump; HP1100 Series Autosampler, HP1100 Series Thermostat, Agilent Ion Trap 1100 SL Mass spectrometer; Working conditions HPLC: Zorbax SB-C18 analytic column 100 mm x 3.0 mmi, d = 0.5 microns.

Mobile Phase: methanol-acetonitrile mixture, 10:90 (v/v), isocratic elution mode; debit: 1mL/min, temperature: 45C, detection MS/MS, mode MRM: injection volume =5 μ L.

Working conditions of MS: ion source: APCI (atmospheric pressure chemical ionization); ionization mode: positive; nebulizer: nitrogen; pressure 60 psi; vaporizer 400C, drying gas: nitrogen; debit 7L/min; temperature = 325C; capillary potential: -4000V.

Analysis mode: monitoring of the sterol specific ions.

Standards: beta-sitosterol, stigmasterol, campesterol and ergosterol

Monitored ions detected through MS methods are presented in Table I. Due to the fact that through ionization all the sterols lose on molecule of water, ions detected by the MS are always in the following form $[M-H_2O+H]^+$.

Received 12th of March 2015. Accepted 20th of April 2015. **Address for correspondence**: Mihaela Orodan, MD, PhD student, "Carol Davila" University of Medicine and Pharmacy Bucharest, Clinical Emergency Hospital Arad, Andrenyi Caroly No.2-4 Street, Arad, Romania; phone/fax: 0040257211233; e-mail: mihaela_noni@yahoo.com

C						
	Sterol	r.t. (min)	М	M-H ₂ O	M-H₂O+H⁺	
	Ergosterol	2.6	396	378	379	
	Brassicasterol	3.3	398	380	381	
	Stigmasterol	4.0	412	394	395	
	Campesterol	4.0	400	382	383	

 Table I. Monitored sterol specific ions in ascending order according to retention time (r.t.)

4.6

Sitosterol

MS detection revealed parent-ions of the analyzed sterols (parent-ions = ions which are formed after ionization before their eventual fragmentation) in each sterol expected ions were revealed (Table I).

414

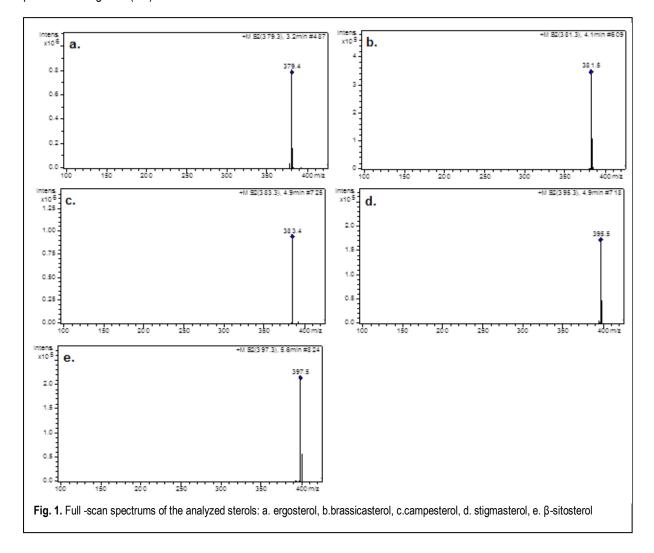
396

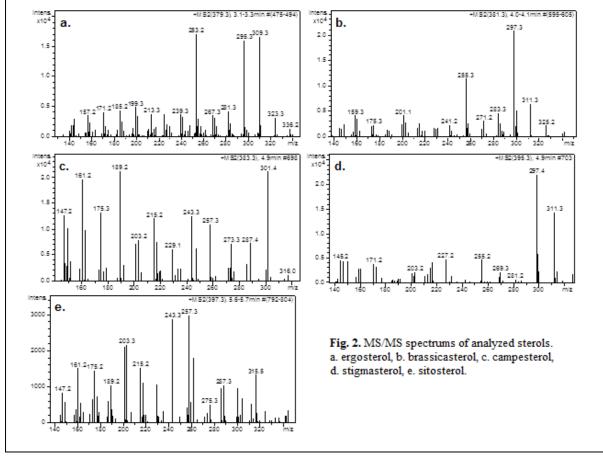
397

Mass spectrum (full-scan type) of the four sterols are presented in Figure 1 (a-e).

This method of analysis (also named MS/MS) is extremely specific as it only registers the intensity of the main ion. An isomer or isobar compound – with the same molecular mass – may give a false positive signal. The analysis on the basis of the fragments from the MS spectrum, which are specific to each structure in turn and which are not the same from one isomer to another, will detect only the compound of interest without interference from others. As well as this, the ion intensity from the MS is proportional with the concentration of the substance in the compound analyzed, so the method may be applied for quantification in place of qualitative analysis.

MS/MS spectrums of the analyzed compounds are presented in Figure 2 (a-e).





MS/MS spectrums of the analyzed compounds are presented in Figure 2 (a-e).

In order to quantify the analyzed sterols, we built a chromatogram for each of the analyzed sterols, according to the intensity of the principle ions from the MS. The ions used for quantification are numbered in Table II.

No.	Compound name	lons specific for identification			
lon [N	Ion [M-H ₂ O+H ⁺] > Ions from spectrum				
1	Ergosterol	379> 253.3,295.3,309.3			
2	Brassicasterol	381>201.3,203.3,215.2,217.3,241.			
		2,255.3,257.4,271.1,297.3,299.3			
3	Campesterol	383>147.3,149.3,161.3,175.3,189.			
		3,203.3,215.3,229.3,243.3,257.3			
2	Stigmasterol	395>163.3,173.2,187.3,199.3,227.			
		2,241.3,255.3,269.2,283.4,297.3			
4	Sitosterol	397>160.9,174.9,188.9,202.9,214.			
		9,243.0,257.0,287.1,315.2			

For the construction of calibration curves, standard sterols were dissolved in acetone (concentration 1 mg/mL), from which successive dilution in acetonitrile were made in order to obtain the calibration solution. The concentration of calibration solution of each analyzed sterol, concentrations found and the accuracy of the determination are presented in Tables III-VII.

 Table III. Concentration of calibration solutions of ergosterol and the accuracy of the determination

Concentration	Ergosterol		
level	Standard concentration (ng/ml)	Concentration found (ng/ml)	Accuracy (%)
1	69.00	70.29	101.87
2	138.00	132.21	95.80
3	276.00	293.67	106.40
4	552.00	514.05	93.12
5	1104.00	1084.81	98.26
6	2208.00	2229.77	100.99
7	3312.00	3534.03	106.70

and the accuracy of their determination					
Concentration	Brassicasterol				
level	Standard	Concentration	•		

found (ng/ml)

60.71

132.74

258.38

489.86

913.66

1071 57

Accuracy %

97.92

107.92

105.03

99.56

92.85

100 10

concentration

(ng/ml) 62.00

123.00

246.00

492.00

984.00

1069 00

1

3

4

5

6

Table IV. Concentration of calibration solutions of brassicasterol

0	1900.00	19/1.3/	100.10
7	2952.00	2931.16	99.29

 $\label{eq:table_table_table} \begin{array}{l} \textbf{Table V}. \ \mbox{Concentration of calibration solution of campesterol} \\ \mbox{and the accuracy of the determination} \end{array}$

Concentration	Campesterol		
level	Standard concentration (ng/ml)	Concentration found (ng/ml)	Accuracy (%)
1	59.00	60.58	102.68
2	117.00	111.11	94.97
3	234.00	235.06	100.45
4	468.00	491.72	105.07
5	936.00	912.98	97.54
6	1872.00	1761.14	94.08
7	2808.00	3046.94	108.51

 Table VI. Concentration of calibration solutions of stigmasterol and their accuracy of determination

Concentration level	Stigmasterol		
	Standard concentration (ng/ml)	Concentration found (ng/ml)	Accuracy (%)
1	136.00	144.6409	106.35
2	272.00	241.2552	88.70
3	544.00	535.5297	98.44
4	1088.00	1152.767	105.95
5	2176.00	2294.101	105.43
6	4352.00	4311.85	99.08
7	6528.00	6656.755	101.97

 Table VII. Concentration of calibration solutions of sitosterol and the accuracy of their determination

Concentration	Sitosterol		
level	standard concentration (ng/mL)	Concentration found (ng/mL)	Accuracy (%)
1	132.00	139.48	105.67
2	264.00	244.58	92.65
3	528.00	528.91	100.17
4	1056.00	1094.38	103.63
5	2112.00	1943.13	92.00
6	4224.00	4250.72	100.63
7	6336.00	6996.62	110.43

A chromatogram of the 5 analyzed sterols in the above conditions, is shown in Figure 3.

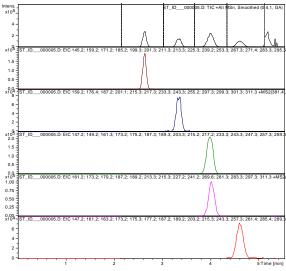
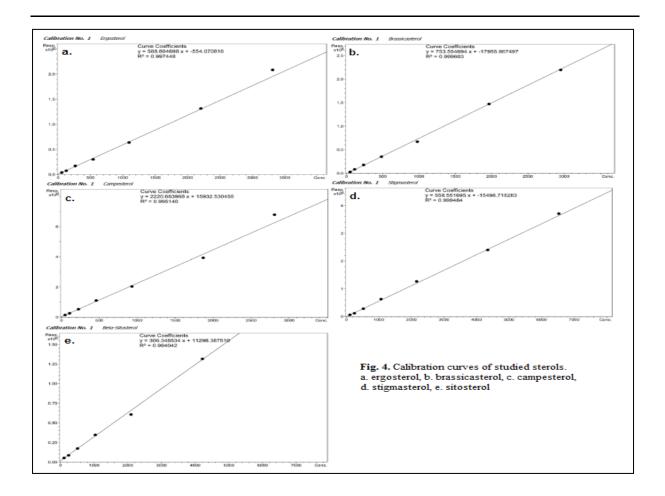


Fig. 3. Chromatograms of the analysed sterols (top to bottom, superimposed signals of standards. a. ergosterol, b. brassicasterol, c. campesterol, d. stigmasterol, e. sitosterol.

Calibration curves of each sterol, on the basis of which the quantification was made are presented in Figure 4 (a-e).



RESULTS AND DISCUSSION

The concentrations of the sterols found are presented in Table VIII. Concentrations according to the dilutions carried out are expressed in Ig/mL.

Table VIII. Stero	concentrations	from analy	zed sterol	S

Sample	Ergosterol	Stigmasterol	Beta-Sitosterol	Campesterol	Brassicasterol
	Concentration (Ig/m	1)			
X1	0.000	0.000	15.859	0.000	0.000
X2	0.000	0.000	17.251	0.000	0.000
Х3	0.000	0.000	21.598	0.000	0.000
X4	0.000	0.000	1.516	0.000	0.000
X5	0.000	0.000	2.033	0.000	0.000
X6	0.000	0.000	0.000	0.000	0.000

Key: X1= RC DH1, X2=BP DH1 , X3=RN DH1, X4= P 30%, X5=VL DH1, X6=CB DH1

Of all the analyzed sterols, the only sterol detected was I-sitosterol. With the exception of the gemmotherapeutic product of *Carpinus betulus* (X6), all samples analyzed contain betasitosterol. The highest concentrations of I-sitosterol are found in the gemmotherapeutic products of the following *Ribes nigrum, Betula pubescens* and *Rosa canina.* Small quantities are found in *Viburnum lantana* and in *propolis tincture.*

CONCLUSIONS

Due to the fact that in our bibliography we didn't find the presence of I-sitosterol, we consider that our research brings a modest contribution to the knowledge of the chemical composition of the gemmotherapeutic products obtained from *Ribes nigrum, Betula pubescens, Rosa canina*.and *Carpinus betulus*.

REFERENCES

Pop G, Galuscan A, Peev C, Militaru A, Vlase L, Ardelean L, Rusu LC. HPLC-MS Identification of Sterol Fractios from Vegetable Oil. *Revista de chimie*, 2012; 63(10): 1046-1050.
 Khalaf I, Vlase L, Ivanescu B, Lazar D, Corciova A. HPLC Analysis of polyphenolic compounds, phytoestrogens and sterols from Glycyrrhiza Glabra L. Tincture. *Studia Ubb Chemia*, 2012; 57(2): 113-118.

3. Khalaf I, Corciovă A, Vlase L, Ivănescu B, Lazăr D. LC/MS Analysis of sterolic compounds from Glycyrrhiza Glabra. *Studia Ubb Chemia*, 2011; 56 (3): 97-102.

ANALIZA COMPUSILOR STEROLICI DIN EXTRACTELE VEGETALE DE RIBES NIGRUM, ROSA CANINA, BETULA PUBESCENS, CARPINUS BETULUS, VIBURNUM LANTANA SI TINCTURA DE PROPOLIS

REZUMAT

Extractele vegetale de tip gemoterapic in dilutie DH1 de *Rosa canina* (X1), *Betula pubescens* (X2), *Ribes nigrum* (X3), *Viburnum lantana* (X5), *Carpinus betulus* (X6) și tinctura 30% de propolis (X4) sunt frecvent folosite în medicina integrativă pentru acțiune antiinflamatoare. Intrucat efectul antiinflamator al unui extract vegetal poate fi imprimat și de steroli, ne-am propus analiza acestora produse (de proveniență Plantextract, Cluj), prin cromatografie de lichide de înalta performanță cuplată cu spectrometrie de masa (LC/MS). S-au analizat: betasitosterolul, stigmasterolul, campesterolul, ergosterolul, brassicasterolul. Concentratiile de steroli gasite, prin metoda mai sus mentionata (exprimate in µg/mL) au fost semnificative doar pentru betasitosterol, în: X1 = 15.859, X2 = 17.251, X3 = 21.598, X4 = 1.516; X5 = 2.033, X6 neidentificabil.

Cuvinte cheie: extracte vegetale de tip gemoterapic, steroli si propolis, analiza LC/MS/MS

ANTIBACTERIAL EFFECTS OF SOME PLANT EXTRACTS ON *STAPHYLOCOCCCUS AUREUS* STRAINS

LAURA M SIMON, LIA PEPELEA, LIA M JUNIE

Microbiology Department, UMF "Iuliu Haţieganu", Cluj-Napoca, Romania

ABSTRACT

Background & Aims: The purpose of this study was to investigate the antibacterial properties of alcoholic extracts and essential oils derived from plants against *S. aureus* strains isolated from patients.

Material and Methods: We studied 50 strains of *S. aureus* isolated from various clinics in Cluj-Napoca, as well as from outpatients and six natural products: Calendula tincture (*Calllendulae flos*), tincture of hawthorn (*Folium Crataegi as flowers*), *Melissa officinalis* essential oil, tincture of sage (*Salviae folium*), propolis tincture and Biosept forte containing 80% thyme essential oil (*Thyme aetheroleum*) and 20% cloves essential oil (*Caryophyllorum aetheroleum*). For each strain susceptibility testing was performed using disk diffusion method on subculture standardized Kirby-Bauer with manual reading.

Results: MSSA strains showed inhibition zone diameters of ≥ 10 mm for *Melissa officinalis* oil at 100% of strains, for propolis tincture in proportion of 93.33% of strains, for Salviae folium at a rate of 86.66% of strains and in 46.66% of strains for both calendula tincture and for hawthorn. MRSA strains showed inhibition zone diameters of ≥ 10 mm, in case of *Melissa officinalis* oil and propolis tincture to 100% of strains, for *Salviae folium* in the proportion of 97.14% of strains, a rate of 28.57% of strains for tincture of hawthorn and the rate of 14.28% of strains for Calendula tincture.

Conclusion: All six plant extracts tested strains of *S. aureus* had antibacterial effects. The best antibacterial effects was observed in case of Biosept forte product. Data showed significant antibacterial effect of *Melissa officinalis* essential oil and propolis. Moderate antibacterial effect was observed for sage tincture (*Salviae folium*), hawthorn (*Folium crataegi*) and marigold (*Calllendulae flos*).

Key words: plant extracts, MRSA, antibacterial effect.

INTRODUCTION

Despite the fact that the pharmaceutical industry produce new antibiotics, in recent years the resistance to chemotherapy is growing. Therefore, there are serious questions over the future use of chemotherapy, which requires taking measures to identify new antibacterial substances.

Currently are evaluated various plant extracts for possible antibacterial effects which covers many bacterial species, including those responsible for causing nosocomial infections, as *Staphylococccus aureus* (*S. aureus*).

The evaluation of antimicrobial activity is based on quantification of bacterial development after the contact with natural extracts.

The purpose of this study was to investigate the antibacterial properties of alcoholic extracts and essential oils derived from plants against *S. aureus* strains isolated from patients.

MATERIALS AND METHODS

We studied 50 strains of *S. aureus* isolated from various clinics in Cluj-Napoca, as well as from outpatients.

The identification of isolates was based on morphological characters, cultural, biochemical and antigenic tests. For each strain susceptibility testing was performed using disk diffusion method on subculture standardized Kirby-Bauer with manual reading (according to CLSI).

The natural products tested were: Calendula tincture (*Calllendulae flos*) from Hofigal, tincture of hawthorn (*Folium Crataegi* as flowers) from Fares, *Melissa officinalis* essential oil from Fares, tincture of sage (*Salviae folium*) from Fares, propolis tincture and Biosept forte (natural antibiotic) from the company Fares containing 80% thyme essential oil (thyme aetheroleum) and 20% cloves essential oil (*Caryophyllorum aetheroleum*). In vitro antibacterial activity of the natural products was tested on solid culture media (Mueller Hinton) distributed in Petri dishes (90 mm).

Received 15th of March 2015. Accepted 20th of April 2015. **Address for correspondence**: Laura M Simon, Microbiology Department, UMF "Iuliu Haţieganu" Cluj-Napoca, 6th Pasteur Street, RO-400349, Cluj-Napoca, Romania; phone/fax: +40264597257, e-mail: lauramihaelasimon@yahoo.com

The preparation of inoculum was made by suspending the colonies in 3 ml saline to the density of a McFarland 0.5 turbidity standard. We used several morphologically similar colonies. The inoculation of the agar plates was made with a sterile cotton swab by removing the excess fluid and by spreading the inoculum over the entire surface of the plate.

After 15 minutes we applied the sterile disks of 5 mm diameter at a distance of 20-25 mm between two neighboring disks. The disks were impregnated with 20µl of each plant extract. We worked with pairs samples, for each strain 2 tests were performed with the same substance. Reading was done after incubation (for 18-20 hours at 37 °C) with a ruler measuring the diameter of the inhibition zones (in mm). We noted the mean (arithmetic average) of the two determinations. We started from the principle that bacteria are more sensitive to the substance studied as the inhibition zone diameter is larger.



Fig. 1. Effects of natural extracts: A (*Calllendulae flos*), B (tincture of hawthorn) (*Folium crataegi* as flowers) and C (*Melissa officinalis* essential oil) on a MSSA strain



Fig. 2. Effects of natural extracts: D (tincture of sage), E (propolis tincture) and F (Biosept forte) on a MSSA strain



Fig. 3. Effects of natural extracts D (tincture of sage), E (propolis tincture) and F (Biosept forte) on *S. aureus* strain ATCC 25923

Statistical methods. To describe qualitative variables we used absolute and relative frequencies plotted by sector and column charts. In case of quantitative variables we used the average as centrality parameter and standard deviation as the dispersion parameter.

RESULTS

In case of MSSA strains, using the tincture of calendula (*Calllendulae flos*), we obtained diameters of the inhibition zones <10 mm for 18% of strains (9 strains) and in case of MRSA strains we recorded values <10 mm were in 62% of strains (31 strains). The inhibition zone diameters between 11 and 20 mm were recorded in 12% of MSSA (6 strains) and 8% MRSA (4 strains) (Figure 4).

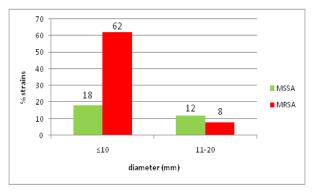


Fig. 4. Diameters of the inhibition zones for Calendula tincture

From 50 strains studied (15 strains MSSA and 35 MRSA strains), 12 strains (24%) showed inhibition zone diameter \ge 10 mm (7 MSSA strains (14%) and 5 MRSA strains (10%)), of which 4 MSSA strains (8%) showed inhibition zone diameter \ge 15 mm (Table I).

Inhibition zone diameter	MSSA	MRSA	TOTAL
≥ 10 mm			%
Calendula tincture			
18	1 (2%)		1(2%)
17	1 (2%)		1(2%)
16	1 (2%)		1(2%)
15	1 (2%)		1(2%)
14	1 (2%)	2 (4%)	3 (6%)
13	-	1 (2%)	1(2%)
11	1 (2%)	1 (2%)	2 (4%)
10	1 (2%)	1 (2%)	2 (4%)
Total	7 (14%)	5 (10%)	12 (24%)

Table I. Diameters of the zones of inhibition ≥ 10 mm for Calendula tincture

We recorded 38 strains (76%) who had the inhibition zone diameters <10 mm, of which 8 MSSA strains (16%) and 30 MRSA strains (60%) (Table II).

 Table II. Diameters of the inhibition zones <10 mm for</th>
 Calendula tincture

Inhibition zone diameter <10 mm Calendula tincture	MSSA	MRSA	TOTAL
Total	8 (16%)	30 (60%)	38 (76%)

Calendula tincture (*Calllendulae flos*) had no effect on 13 strains (26%): 9 MRSA strains (18%) and 4 MSSA strains (8%). Thus, Calendula tincture showed antibacterial effect of the 12 strains (24%) of the 50 strains tested. *Calendula* tincture (*Calllendulae flos*) had no effect on the 38 strains (76%), 30 MRSA strains (60%) and 8 MSSA strains (16%). For hawthorn tincture (*Folium crataegi*) the values of the diameters of the inhibition zones for MSSA were less than 10 mm in 18% of strains (9 strains) and in in 60% of MRSA strains (30 strains). The diameters of the of inhibition zones were between 11 and 20 mm in 12% of MSSA strains (6 strains) and in 10% of MRSA strains (5 strains) (Figure 5).

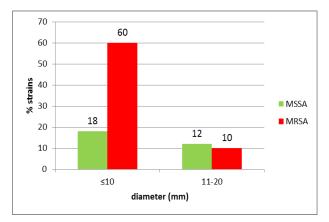


Fig. 5. Diameters of the inhibition zones for hawthorn tincture

From all studied strains, 17 strains (34%) showed an inhibition zone diameter \geq 10 mm: 7 MSSA strains (14%) and 10 MRSA strains (20%). 3 MSSA strains (6%) showed inhibition zone diameters \geq 15 mm (Table III).

Table III. Diameters of the inhibition zones ≥ 10 mm for Hawthorn tincture

Inhibition zone diameter ≥ 10 mm Hawthorn tincture (<i>Folium crataegi</i>)	MSSA	MRSA	TOTAL
18	-		-
17	1 (2%)		1 (2%)
16	1 (2%)		-
15	1 (2%)		1(2%)
14	2 (4%)	4 (6%)	6 (12%)
13	1 (2%)	1 (2%)	2(4%)
12	-	-	
11	-	-	-
10	1 (2%)	5 (10%)	6 (12%)
Total	7 (14%)	10 (20%)	17 (34%)

Thus, the hawthorn tincture showed antibacterial effect on 17 strains (34%) of the 50 tested strains and had no effect on 33 strains(66%): 25 MRSA strains (50%) and 8 MSSA strains (16%).

For *Melissa officinalis* essential oil, the diameters of the inhibition zones for MSSA ranged from 21-40 mm in 12% of strains (6 strains) and they were less than 20 mm in 18% of strains (9 strains). In case of MRSA inhibition zone diameter values were less than 20 mm in 64% of strains (32 strains) and were between 21 and 40 mm in 6 strains (Figure 6).

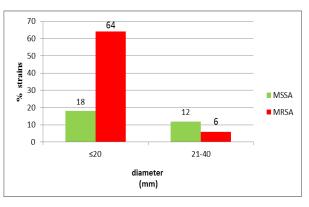


Fig. 6. Diameters of the inhibition zones for *Melissa officinalis* essential oil

In case of sage tincture (*Salviae folium*), the diameters of the inhibition zones were for MSSA strains between 9 and 27 mm, and for MRSA strains they were between 8-18 mm. 9 strains were resistant. Diameters ranging from 15 to 18 mm have been reported to 6 MSSA strains (12%) and to 14 MRSA strains (28%) (Figure 7).

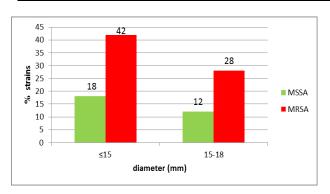


Fig. 7. Diameters of the inhibition zones for sage tincture

For propolis tincture, the diameters of the inhibition zones for MSSA strains were between 9-26 mm. For MRSA strains, the diameters were between 16 and 24 mm. Diameters between 21 and 26 mm have been reported for 7 strains of MRSA (14%) and 6 MSSA strains (12%) (Figure 8).

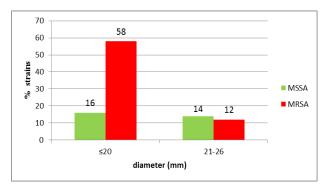


Fig. 8. Diameters of the inhibition zones for propolis tincture

In case of Biosept forte, the diameters of the zones of inhibition for MSSA were between 51-61 mm. For MRSA the diameters of the zones of inhibition ranged between 40-65 mm. We reported diameters larger than 51 mm at 12 MSSA strains (24%) and at 28 MRSA strains (56%) (Figure 9).

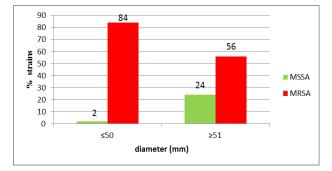


Fig. 9. Diameters of the zones of inhibition for Biosept forte

MSSA strains showed inhibition zone diameters of \geq 10 mm for *Melissa officinalis* oil at 100% of strains, for

propolis tincture in proportion of 93,33% of strains, for *Salviae folium* at a rate of 86,66% of strains and in 46,66% of strains for both *Calendula* tincture and for hawthorn.

MRSA strains showed inhibition zone diameters of \geq 10 mm, in case of *Melissa officinalis* oil and propolis tincture to 100% of strains, for *Salviae folium* in the proportion of 97,14% of strains, a rate of 28,57% of strains for tincture of hawthorn and the rate of 14,28% of strains for *Calendula* tincture (Figure 10).

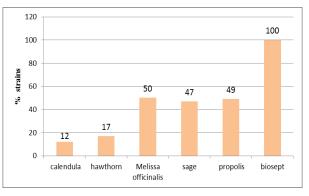


Fig. 10. Diameters of the inhibition zones \ge 10 mm for MSSA and MRSA strains

DISCUSSION

Given the alarming rise of antibiotic resistance, there is a growing concern among researchers for the discovery of new antibacterial substances. Thus, the study of antibacterial effects of plant extracts is very important. The use of plants in therapy has many advantages, including: low cost and low toxicity when administered at high doses.

Plants contain components with antibacterial activity whose use is increasing worldwide. Antioxidants are considered protective agent, serving to enhance the oxidative processes. The components with antioxidant effects are phenolic compounds, especially flavonoids, which interact with free radicals. Antibacterial effect could be the result of induction of free radicals [6,10].

In our study, the most intense antimicrobial product can be considered Biosept forte, which presented inhibition zone diameters of 65 mm maximum, followed by essential oil of *Melissa officinalis* with maximum 40 mm inhibition zone diameters.

Propolis tincture had also an important antimicrobial effect, for which the maximum diameter of the inhibition zones was 26 mm and 28% from all strains presented a diameter between 21 and 26 mm.

In case of these 50 strains tested strains, the inhibition zone diameters for sage tincture ranged between 15 and 18 mm for 40% of strains, showing an antimicrobial effect.

Hawthorn tincture (*Folium crataegi*) had no effect on 78% of *S. aureus* strains tested, and in case of marigold tincture (*Calllendulae flos*), 80% of strains were found to be resistant.

Based on these diameters we can conclude that most plant extracts showed antibacterial effect.

A recent study in Brazil reveals anti-inflammatory, antibacterial and angiogenic properties of *C. officinalis* [14].

A study based on several aerobic and facultative anaerobic bacterial species including S. aureus indicates a moderate inhibitory activity with MIC> 2048 mg / L in all bacteria tested, except *Prevotella spp* [9]. In Portugal, a study on several herbs highlights in case of *Crataegus monogyna* a highest concentration of phenolic acids (5.5 mg/gdw), which proves an important antioxidant activity [2].

Melissa officinalis has sedative [20], antiviral and antibacterial effects [5]. Good antimicrobial effects were recorded on a multidrug-resistant strains of *Shigella sonei*.

Antimicrobial effects of sage is due to its components: oleanolic acid, triterpenoid and were identified MRSA and *Streptococcus pneumoniae* [8,22].

A study on Salvia corrugator Vahl identified antibacterial activity on Gram-positive bacteria of two components: fruticuline A and demetilfruticuline. The first presented bacteriostatic activity on all tested strains and the two showed significant bactericidal activity against *S. aureus* and *S. epidermidis* and bacteriostatic activity on *Enterococcus faecalis* and *Enterococcus faecium* [4].

A study made in Tunisia shows antimicrobial and antioxidant effect of methanolic extracts of sage [16], while another study in Jordan on several plant extracts, including sage showed no antibacterial effect on *S. aureus* and *P. aeruginosa* [1].

Takaisi & Schilcher have suggested that the antimicrobial action of propolis is due to the combined action of some constituents that inhibit the bacterial RNA polymerase (pinocembrine, galangine and caffeic acid–phenethyl-ester) [21].

There are studies that show the antibacterial effect of propolis on MRSA [3,15], it is higher than the antibacterial effect of honey [13] and more pronounced on Gram-positive [18] bacteria compared to gram-negative bacteria and fungi [19]. Propolis increases the antibacterial effect of antibiotics such as Ampicillin, Gentamicin, Netilmicin, Chloramphenicol, Vancomycin [7] and Streptomycin [17]. The antibacterial effect against *S. aureus* was evidentiated in Japan, but data obtained emphasises its variation depending on the area and the month in which this was collected [12].

The essential oil of *Thymus vulgaris* showed an antibacterial effect on *E. coli* [11].

CONCLUSIONS

- All six plant extracts tested strains of *S. aureus* had antibacterial effects;
- The best antibacterial effects was observed in case of Biosept forte product;
- Data showed significant antibacterial effect of Melissa officinalis essential oil and propolis;
- Moderate antibacterial effect was observed for sage tincture (*Salviae folium*), hawthorn (*Folium crataegi*) and marigold (*Calllendulae flos*).

REFERENCES

1. Abu-Darwish MS, Al-Ramamneh EA, Kyslychenko VS, *et al.* The antimicrobial activity of essential oils and extracts of some medicinal plants grown in Ash-shoubak region - South of Jordan. *Pak J Pharm Sci* 2012; 25(1): 239-46.

2. Barros L, Dueñas M, Carvalho AM, *et al.* Characterization of phenolic compounds in flowers of wild medicinal plants from Northeastern Portugal. *Food Chem Toxicol* 2012; 50(5):1576-82.

3. Berretta AA, Nascimento AP, Bueno PC, *et al.* Standardized Extract (EPP-AF®), an Innovative Chemically and Biologically Reproducible Pharmaceutical Compound for Treating Wounds. *Int J Biol Sci* 2012; 8(4): 512-21.

4. Bisio A, Romussi G, Russo E, *et al.* Antimicrobial activity of the ornamental species Salvia corrugata, a potential new crop for extractive purposes. *J Agric Food Chem* 2008; 56(22): 10468-72.

5. Canadanović-Brunet J, Cetković G, Djilas S, *et al.* Radical scavenging, antibacterial, and antiproliferative activities of Melissa officinalis L. extracts. *J Med Food* 2008; 11(1): 133-43.

6. Dawidowicz, A.L., Wianowska D, Baraniak B. The antioxidant properties of alcoholic extracts from Sambucus nigra L (antioxidant properties of extracts). *LWT-Food Sci. Technol* 2006; 39: 308-15.

7. Fernandes Júnior A, Balestrin EC, *et al.* Propolis: anti-Staphylococcus aureus activity and synergism with antimicrobial drugs. *Mem Inst Oswaldo Cruz* 2005; 100(5): 563-6.

8. Horiuchi K, Shiota S, Hatano T, *et al*. Antimicrobial activity of oleanolic acid from Salvia officinalis and related compounds on vancomycin-resistant enterococci (VRE). *Biol Pharm Bull* 2007; 30(6): 1147-9.

9. lauk L, Lo Bue AM, Milazzo I, *et al.* Antibacterial activity of medicinal plant extracts against periodontopathic bacteria. *Phytother Res* 2003; 17(6): 599-604.

10. Inci E, Civelek S, Seven A, *et al.* Laryngeal cancer: In relation with oxidative stress. *Tohoku J Exp Med* 2003; 200: 17-23.

11. Jugl-Chizzola M, Spergser J, Schilcher F, *et al.* Effects of Thymus vulgaris L. as feed additive in piglets and against haemolytic E. coli in vitro. *Berl Munch Tierarztl Wochenschr* 2005; 118: 495-501.

12. Lu LC, Chen YW, Chou CC. Antibacterial activity of propolis against Staphylococcus aureus. *Int J Food Microbiol* 2005; 102(2): 213-20.

13. Miorin PL, Levy Junior NC, Custodio AR, et al. Antibacterial activity of honey and propolis from Apis mellifera and

Tetragonisca angustula against Staphylococcus aureus. *J Appl Microbiol* 2003; 95(5): 913-20.

14. Parente LM, Lino Júnior RS, Tresvenzol LM, *et al.* Wound Healing and Anti-Inflammatory Effect in Animal Models of Calendula officinalis L. Growing in Brazil. *Evid Based Complement Alternat Med* 2012: 375671.

15. Raghukumar R, Vali L, Watson D, Fearnley J, Seidel V. Antimethicillin-resistant Staphylococcus aureus (MRSA) activity of 'pacific propolis' and isolated prenylflavanones. *Phytother Res* 2010; 24(8): 1181-7.

16. Salah KB, Mahjoub MA, Ammar S, *et al.* Antimicrobial and antioxidant activities of the methanolic extracts of three Salvia species from Tunisia. *Nat Prod Res* 2006; 20(12): 1110-20.

17. Scazzocchio F, D'Auria FD, Alessandrini D, *et al.* Multifactorial aspects of antimicrobial activity of propolis. *Microbiol Res* 2006; 161(4): 327-33.

18. Seidel V, Peyfoon E, Watson DG, et al. Comparative study of the antibacterial activity of propolis from different

geographical and climatic zones. *Phytother Res* 2008; 22(9): 1256-63.

19. Silici S, Kutluca S. Chemical composition and antibacterial activity of propolis collected by three different races of honeybees in the same region. *J Ethnopharmacol* 2005; 99(1): 69-73.

20. de Sousa AC, Alviano DS, Blank AF, *et al*. Melissa officinalis L. essential oil: antitumoral and antioxidant activities. *J Pharm Pharmacol* 2004; 56(5): 677-81.

21. Takaisi NB, Schilcher H. Electron microscopy and microcalorimetric investigations of the possible mechanism of the antibacterial action of a defined propole provenance. *Planta Med* 1994; 60: 222-7.

22. Weckesser S, Engel K, Simon-Haarhaus B, *et al.* Screening of plant extracts for antimicrobial activity against bacteria and yeasts with dermatological relevance. *Phytomedicine* 2007; 14(7-8): 508-16.

EFECTELE ANTIBACTERIENE ALE UNOR EXTRACTE DIN PLANTE ASUPRA TULPINILOR DE *STAPHYLOCOCCCUS AUREUS*

REZUMAT

Scop: Scopul acestui studiu a fost de a investiga proprietatile antibacteriene ale extractelor alcolice si uleiurilor esentiale derivate din plante asupra tulpinilor de *S. aureus* isolate de la pacienti.

Materiale si metode: Am studiat 50 de tulpini de *S. aureus* isolate din diferite institutii spitalicesti din Cluj-Napoca, precum si de la pacienti tratati ambulator, si 6 produse naturale: tinctura de galbenele (*Calllendulae flos*), tinctura de paducel (*Folium Crataegi as flowers*), ulei esential de *Melissa officinalis*, tinctura de salvie (*Salviae folium*), tinctura de propolis si Biosept forte, care contine 80% ulei esential de cimbru (*Thyme aetheroleum*) si 20% ulei esential de cuisoare (*Caryophyllorum aetheroleum*). Pentru fiecare tulpina, testarea suscetibilitatii a fost efectuata folosind metoda difuzimetrica standardizata a subculturilor Kirby-Bauer cu citire manuala.

Rezultate: Tulpinile de MSSA au prezentat zone de inhibitie cu diametrul \ge 10 mm la uleiul de *Melissa officinalis* pentru 100% dintre tulpini, la tinctura de propolis in proportie de 93,33% dintre tulpini, la *Salviae folium* 86,66% dintre tulpini si 46,66% dintre tulpini la tinctura de galbenele si paducel. Tulpinile de MRSA au prezentat zone de inhibitie cu diametrul \ge 10 mm in cazul uleiului de *Melissa officinalis* si a tincturii de propolis pentru 100% dintre tulpini, 97,14% tulpini pentru *Salviae folium*, 28,57% tulpini pentru tinctura paducel si 14,28% tulpini pentru tinctura de galbenele.

Concluzii: Toate cele sase extracte din plante testate pe tulpinile de *S. aureus* au prezentat efecte antibacteriene. Cele mai bune efecte antibacteriene au fost observate in cazul produsului Biosept forte. Datele arata un effect antibacterian semnificativ al uleiului esential de *Melissa officinalis* si propolis. Efect antibacterian moderat a fost observat in cazul tincturii de salvie (*Salviae folium*), paducel (*Folium crataegi*) si galbenele (*Calllendulae flos*). **Cuvinte cheie:** extracte din plante, plant extracts, MRSA, efect antibacterian.

ACUTE ORAL AND DERMAL EFFECTS OF SPIRULINA IN MICE

FLORINA-MARIA ANDRICA¹, CRISTINA DEHELEAN², MARIA-CORINA SERBAN^{3*}, IULIA PÂNZARU², DORINA CORICOVAC², SIMONA DRĂGAN⁴

¹Department of Pharmaceutical Chemistry, Faculty of Pharmacy

²Department of Toxicology, Faculty of Pharmacy

³Department of Functional Sciences, Faculty of Medicine

⁴Department of Cardiology, Faculty of Medicine

"Victor Babes" University of Medicine and Pharmacy, Timisoara

ABSTRACT

Spirulina (SP) is a spiral-shaped, blue-green microalga included in bacteria kingdom. SP has a rich content in essential amino acids, phycobiliproteins (phycoerythrin, phycocyanin), minerals, vitamins, carotenoids, polyphenols and essential fatty acids. The aim of this study consists in a safety evaluation of SP treatment using oral and dermal toxicity studies. SP has revealed no significant changes in skin parameters, behavior or other locomotion problems when was administered at limited doses of 2000 mg/bw/24 h and 5000 mg/bw/24 h on SKH1 mice. Consequently, SP is a natural product that can be used both by adults and children, being characterized by multi-target beneficial effects for all whole human body and, nevertheless, by lack of toxicity.

Keywords: spirulina, acute toxicity, skin toxicity, subchronic toxicity, oral toxicity

INTRODUCTION

Spirulina (SP) is a spiral-shaped, blue-green alga [1] which belongs to the Oscillateriaceae family [2]. Firstly, SP was included in the plant kingdom, although, in the present it belongs to the bacteria kingdom because of its biochemical, physiological and genetics properties [3]. The most commonly used species of SP are: Arthrospira platensis, Arthrospira maxima and Arthrospira fusiformis [3,4]. SP has a rich content in essential amino acids, phycobiliproteins (red phycoerythrin, blue phycocyanin) [5], minerals, vitamins (B₁, B₂, B₃, B₅, B₆, B₇, B₉, B₁₂, D, E), carotenoids (alpha-carotene, beta-carotene, zeaxanthin) [6,7] and essential fatty acids (3.6 gamma-linolenic acid, linoleic acid, stearidonic acid and arachidonic acid) [8-12]. The methanolic extract of SP has also revealed high amounts of phenolic compounds including salicylic, trans-cinnamic, chlorogenic and caffeic acids that have been responsible for its antioxidant effects [13].

Furthermore, the microelements of SP depend both on the type and the mineral pollution [14] of producing areas [15]. SP and its derivatives are widely used for their antioxidant, anticancer, anti-inflammatory, anti-diabetic, antimicrobial, as well as for their immunostimulant effects [16-18]. In this regard, SP is generally recommended in various chronic pathologies including arterial hypertension, hypercholesterolemia, malnutrition, obesity, sugar diabetes and cancer [19,20]. SP is considered to be one of the most complex natural products that is widely used not only by adults but also by children [21]. The aim of this study consists in a safety evaluation of SP treatment using oral and dermal toxicity studies.

MATERIALS AND METHODS

This experimental study was done on healthy young adult SKH1 mice. The females were non-pregnant. All the animals involved in the study were between 8 and 12 weeks old. The mice were housed in standard conditions: room temperature 22°C (+ 3°C), relative humidity between 30% and 70 % and artificial lighting characterized by 12 hours of light and 12 hours dark. During the experiment the mice have received food consisting in special pellets and water *ad libitum*. The animals were randomly selected, marked for individual identification and acclimated to the laboratory conditions for 5 days before to begin the experiment. The animals that were included in the study have appropriate size and age [22].

Spirulina, a dark blue-green powder was purchased from FAVISAN Laboratories, Lugoj, Romania.

Received 16th of May 2015. Accepted 10th of June 2015. **Address for correspondence**: Lecturer Maria-Corina Serban, MD, PhD, MSc, Department of Functional Sciences, University of Medicine and Pharmacy "Victor Babes" Timisoara, Eftimie Murgu Square No. 2A, RO-300041, Timisoara, Romania; phone: +40752444900, fax: +40256220479; e-mail: dr.corinaserban@vahoo.com

Acute toxicity studies

Hairless SKH1 mice weighing 25 - 30 g were included in the study. Acute oral and dermal toxicities studied were performed according to OECD- 425 and OECD-434 guidelines. Because SP extract is widely used for centuries all around the world with no major side effects we concluded to apply the limit test for evaluating the acute oral and dermal toxicity of this natural product. According to OECD-425, only one mouse was firstly selected for the acute oral study. It was fasted overnight and in the next morning it was dosed with a test dose of 2000 mg/bw/24h. The animal has survived, and we continued to dose other four female mice with the same dose of SP. The animals have also survived. After that, we continued to give a dose of 5000 mg/bw/24 h to one mouse firstly and after that to the other three mice. The animals were observed for side effects for 14 days and after that were sacrificed.

According to OECD-434, for acute dermal toxicity test were selected 18 mice having approximately the same age and weight. These mice were divided in 4 groups: 5 female mice (group 1), 5 male mice (group 2), 4 healthy mice treated with diluted solution of alcohol (2 males, 2 females) (group 3), 4 healthy untreated mice (2 males, 2 females). SP was dissolved in a diluted alcoholic solution (67%) and, then, was applied uniformly over 10 % of the total body surface area. The skin parameters (melanin levels and erythema) were decelated using a mexameter MX18 (Courage& Khazaka Electronics, Germany) and the skin hydratation was evaluated using a corneometer CM825 MX18 (Courage&Khazaka Electronics, Germany). All animals were observed for 14 days after application of SP extract and after that were sacrificed.

The experimental study protocol was designed according to the Universal Declaration of Animal Rights proclaimed in Paris in 1978 and to the Declaration of Helsinki, the European Convention (ETS No. 123) amended in 1998 (ETS No. 170) and Council Directive 86/609/EEC on the protection of vertebrate animals used for experimental and other scientific purposes. The animal study protocol was approved by the Ethics Committee of the Faculty of Pharmacy, "Victor Babes" University of Medicine and Pharmacy, Timisoara, Romania.

RESULTS

It was observed that all the mice that received SP in both doses of 2000 mg/bw/24 h and 5000 mg/bw/24 h have survived and they did not manifest any behavioral changes, convulsions, locomotion problems or mortality (Figure 1). Consequently, the LD50 for SP was greater than 5000mg/bw/24h.

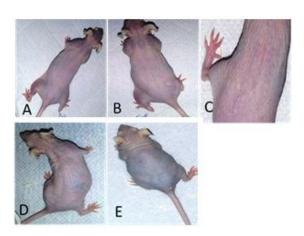


Fig. 1. Mice – 24 hours after the skin application of SP extract: A) male mice treated with alcoholic solution of SP; B) female mice treated with alcoholic solution of SP; C) skin of female mice treated with alcoholic solution of SP; D) female mice treated with alcoholic solution; E) female mice untreated

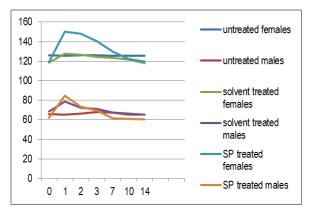


Fig. 2. Evolution of melanin levels in mice

There were no significant differences between the values of melanin before and after SP application in female mice (p = 0.2097) p>0.05 and in male mice (p = 0.7517) p>0.05.

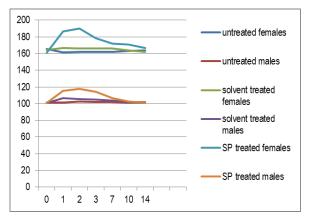


Fig. 3. Evolution of erythema in mice

The erythema values increased in the first 48 h after local administration of SP both in female (p = 0.035) p<0.05 and male mice (p = 0.05222) p>0.05 but significant changes were seen only in female mice treated with SP. The alcoholic solution that was used as solvent did not caused significant differences in female (p = 0.1000) p>0.05 and in male groups (p = 0.071) p>0.05. Consequently, the significant changes in females group treated with SP were caused only by SP and not by the solvent.

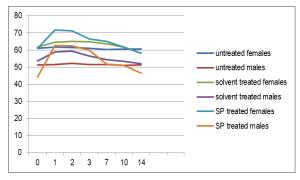


Fig. 4. Evolution of skin hydratation levels in mice

There were no significant differences regarding the skin hydratation measurements before and after the local application of SP in female (p = 0.060) and in male group (p = 0.3830), both p > 0.05. Consequently, according to our toxicity studies on mice, we concluded that SP has lack oral and dermal toxicity, because the majority of the skin parameters that were measured after the local administration of SP extract did not suffer significant changes. The erythema levels were significantly higher only in the female group treated with SP. However, the information regarding the side effects of SP treatments is still limited. According to another experimental study, higher dose of SP (1000 mg/kg) did not cause important changes in cardiac parameters in mice [23].

Some clinical trials tested the toxicity of SP in humans. There have been reported only minor hypersensitivity reactions including urticarial, rash, diaphoresis and bronchospasm [24]. Other side effects observed in patients treated daily with 1g of SP were: flushing of the face, headache, muscle pain, sweating and difficulty concentrating [25]. Moreover, it was reported a higher rate of protidemia and creatinemia in HIV patients treated with SP for 6 month [26]. It has been reported a clinical case of an 82-year-old healthy woman who manifested bullae, partly hemorrhagic, on the trunk and extremities, secreting erosions and submammary macerations after a 2-year period of SP intake. Another 57-year-old man diagnosed with pemphigus vulgaris has developed a severe flare-up of the disease after consumption of a supplements containing Ginseng, Ginkgo biloba and SP. Another case of a 45-year-old woman who presented dermatomyositis after intake of a supplement based on methylsulfonylmethane, organic cayenne pepper, the algae Aphanizomenon flos-aquae and SP was described [27]. Moreover, SP caused anaphylaxis to a 17-year-old male after one ingestion of a SP tablet [28] and acute rhabdomyolysis to another patient which consumed a dietary supplement containing SP [29].

According to Health Food Organization, 4g of SP is daily recommended for an adult of 50 kg body weight [30]. The beneficial effects of SP are recognized by World Health Organization (WHO), Food and Agriculture Organization of the United Nations, United Nations Industrial Development Organization and United Nations Children's Fund [2]. The WHO projects considered SP to be one of the most curative and prophylactic components of nutrition in the 21st century [31] due to its remarkable nutrient profile, lack of toxicity and therapeutic effects [32]. The Dietary Supplements Information Expert Committee of the United States Pharmacopeia Convention assigned a class A safety rating for SP maxima and SP platensis, including these dietary supplement ingredients as monographs in United States Pharmacopeia and National Formulary [33].

CONCLUSION

The acute oral and dermal toxicity studies that were made on hairless SKH1 mice using limit doses of 2000 mg/bw/24 h and 5000 mg/bw/24 h of SP revealed no significant changes in skin parameters, behavior or other locomotion problems. Moreover, all the animals included in study have survived; in female group it has been observed a higher level of erythema parameters. We consider that SP is a complex natural product with multi-target beneficial effects in rats characterized by lack of toxicity that can be used both by adults and children.

COMPETING INTERESTS

The authors declare that they have no competing interests.

ACKNOWLEDGEMENT

This work was supported by the POSDRU grant no. 159/1.5/S/136893: "Strategic partnership for the increase of the scientific research quality in medical universities through the award of doctoral and postdoctoral fellowships – DocMed.Net_2.0"

REFERENCES

1. Hongsthong A, Sirijuntarut M, Prommeenate P, Thammathorn S, Bunnag B, Cheevadhanarak S, et al. Revealing differentially expressed proteins in two morphological

forms of Spirulina platensis by proteomic analysis. *Molecular Biotechnology.* 2007; 36(2): 123-30.

2. Choi WY, Kang do H, Lee HY. Enhancement of Immune Activation Activities of Spirulina maxima Grown in Deep-Sea Water. *International Journal of Molecular Sciences*. 2013; 14(6): 12205-21.

3. Deng R, Chow TJ. Hypolipidemic, antioxidant, and antiinflammatory activities of microalgae Spirulina. *Cardiovascular Therapeutics*. 2010; 28(4): e33-45.

4. Thengodkar RR, Sivakami S. Degradation of Chlorpyrifos by an alkaline phosphatase from the cyanobacterium Spirulina platensis. *Biodegradation*. 2010; 21(4): 637-44.

5. Seo YC, Choi WS, Park JH, Park JO, Jung KH, Lee HY. Stable Isolation of Phycocyanin from Spirulina platensis Associated with High-Pressure Extraction Process. *International Journal of Molecular Sciences*. 2013; 14(1): 1778-87.

6. Choi WY, Kang DH, Lee HY. Enhancement of immune activation activities of Spirulina maxima grown in deep-sea water. *International Journal of Molecular Sciences*. 2013; 14(6): 12205-21.

7. Akao Y, Ebihara T, Masuda H, Saeki Y, Akazawa T, Hazeki K, et al. Enhancement of antitumor natural killer cell activation by orally administered Spirulina extract in mice. *Cancer Science.* 2009; 100(8): 1494-501.

8. Grawish M, Zaher A, Gaafar A, Nasif W. Long-term effect of Spirulina platensis extract on DMBA-induced hamster buccal pouch carcinogenesis (immunohistochemical study). *Medical Oncology.* 2010; 27(1): 20-8.

9. Ogato T, Kifle D, Fetahi T, Sitotaw B. Evaluation of growth and biomass production of Arthrospira (Spirulina) fusiformis in laboratory cultures using waters from the Ethiopian soda lakes Chitu and Shala. *Journal of Applied Phycology.* 2014; 26(6): 2273-82.

10. Nagaoka S, Shimizu K, Kaneko H, Shibayama F, Morikawa K, Kanamaru Y, et al. A novel protein C-phycocyanin plays a crucial role in the hypocholesterolemic action of Spirulina platensis concentrate in rats. *The Journal of Nutrition*. 2005; 135(10): 2425-30.

11. Thaakur SR, Jyothi B. Effect of spirulina maxima on the haloperidol induced tardive dyskinesia and oxidative stress in rats. *Journal of Neural Transmission*. 2007; 114(9): 1217-25.

12. Cheng CG, Hong QH, Li DT, Fan MH, Cai XD. Determination of trace elements in Spirulina platensis (Notdst.) Geitl. by flame atomic absorption spectrometry combined with microsampling pulse nebulization technique. *Guang pu xue yu guang pu fen xi* = *Guang pu*. 2006; 26(9): 1735-7.

13. Miranda MS, Cintra RG, Barros SBM, Mancini-Filho J. Antioxidant activity of the microalga Spirulina maxima. *Brazilian Journal of Medical and Biological Research*. 1998; 31: 1075-9.

14. Vicat JP, Doumnang Mbaigane JC, Bellion Y. Contents of macromineral and trace elements in spirulina (Arthrospira platensis) from France, Chad, Togo, Niger, Mali, Burkina-Faso and Central African Republic. *Comptes rendus biologies*. 2014; 337(1): 44-52.

15. Guan Y, Zhao HY, Ding XF, Zhu YY. Analysis of the contents of elements in spirulina from different producing areas. *Guang pu xue yu guang pu fen xi* = *Guang pu.* 2007; 27(5): 1029-31.

16. Hosseini SM, Khosravi-Darani K, Mozafari MR. Nutritional and medical applications of spirulina microalgae. *Mini Reviews in Medicinal Chemistry*. 2013; 13(8): 1231-7.

17. Kulshreshtha A, Zacharia AJ, Jarouliya U, Bhadauriya P, Prasad GB, Bisen PS. Spirulina in health care management. *Current Pharmaceutical Biotechnology*. 2008; 9(5): 400-5.

18. Araldi RP, Rechiutti BM, Mendes TB, Ito ÉT, Souza EB. Mutagenic potential of Cordia ecalyculata alone and in association with Spirulina maxima for their evaluation as candidate anti-obesity drugs. *Genetics and Molecular Research*. 2014; 13(3): 5207-20.

19. Duan M, Ma WX, Li L, Sun XT. Determination of micro-elements in natural spirulina using FAAS. *Guang pu xue yu guang pu fen xi* = *Guang pu*. 2001; 21(6): 868-70.

20. Moura LP, Puga GM, Beck WR, Teixeira IP, Ghezzi AC, Silva GA, et al. Exercise and spirulina control non-alcoholic hepatic steatosis and lipid profile in diabetic Wistar rats. *Lipids in Health and Disease.* 2011; 10:77.

21. Al-Dhabi NA. Heavy metal analysis in commercial Spirulina products for human consumption. *Saudi Journal of Biological Sciences*. 2013; 20(4): 383-8.

22. Sudha S, Karthic R, Naveen JR. Anti hyperlipidemic activity of Spirulina platensis in Triton x-100 induced hyperlipidemic rats. *Hygeria JD Med.* 2011; 3(2): 32-7.

23. Ibrahim AE, Abdel-Daim MM. Modulating Effects of Spirulina platensis against Tilmicosin-Induced Cardiotoxicity in Mice. *Cell J.* 2015; 17(1): 137-44.

24. Haller C, Keamey T, Bent S, Ko R, Benowitz N, Olson K. Dietary supplement adverse events: report of a one-year poison center surveillance project. *Journal of Medical Toxicology.* 2008; 4(2): 84-92.

25. Ravi M, De SL, Azharuddin S, Paul SF. The beneficial effects of Spirulina focusing on its immunomodulatory and antioxidant properties. *Nutr Diet Suppl.* 2010; 2: 73-83.

26. Yamani E, Kaba-Mebri J, Mouala C, Gresenguet G, Rey JL. Use of spirulina supplement for nutritional management of HIV-infected patients: study in Bangui, Central African Republic. *Medecine Tropicale: Revue du Corps de Sante Colonial.* 2009; 69(1): 66-70.

27. Kraigher O, Wohl Y, Gat A, Brenner S. A mixed immunoblistering disorder exhibiting features of bullous pemphigoid and pemphigus foliaceus associated with Spirulina algae intake. *International Journal of Dermatology*. 2008; 47(1): 61-3.

28. Le TM, Knulst AC, Rockmann H. Anaphylaxis to Spirulina confirmed by skin prick test with ingredients of Spirulina tablets. *Food and Chemical Toxicology*. 2014; 74: 309-10.

29. Mazokopakis EE, Karefilakis CM, Tsartsalis AN, Milkas AN, Ganotakis ES. Acute rhabdomyolysis caused by Spirulina (Arthrospira platensis). *Phytomedicine*. 2008; 15(6-7): 525-7.

30. Ishimi Y, Sugiyama F, Ezaki J, Fujioka M, Wu J. Effects of spirulina, a blue-green alga, on bone metabolism in ovariectomized rats and hindlimb-unloaded mice. *Bioscience, Biotechnology, and Biochemistry.* 2006; 70(2): 363-8.

31. Marcel AK, Ekali LG, Eugene S, Arnold OE, Sandrine ED, von der Weid D, et al. The effect of Spirulina platensis versus soybean on insulin resistance in HIV-infected patients: a randomized pilot study. *Nutrients.* 2011; 3(7): 712-24.

32. Savranoglu S, Tumer TB. Inhibitory effects of spirulina platensis on carcinogen-activating cytochrome P450 isozymes and potential for drug interactions. *International Journal of Toxicology*. 2013; 32(5): 376-84.

33. Marles RJ, Barrett ML, Barnes J, Chavez ML, Gardiner P, Ko R, et al. United States pharmacopeia safety evaluation of spirulina. *Critical Reviews in Food Science and Nutrition.* 2011; 51(7): 593-604.

EFECTELE ACUTE ORALE ȘI CUTANATE ALE SPIRULINEI PE MODEL ANIMAL DE ȘOARECE

REZUMAT

Spirulina (SP) este o microalgă albastră-verde, cu formă spiralată, fiind inclusă în regnul bacteriilor. SP are un conţinut bogat în aminoacizi esenţiali, ficobiliproteine (ficoeritrină şi ficocianină), minerale, vitamine, carotenoizi, polifenoli şi acizi graşi esenţiali. Scopul acestui studiu constă în evaluarea siguranţei tratamentului cu SP prin aplicarea unor texte de toxicitate acută orală şi cutanată. În urma tratamentului oral şi local cu SP utilizându-se doze limită de 2000 mg/kg corp/24h şi 5000 mg/kg corp/24 h nu s-au observat modificări semnificative în ceea ce priveşte parametrii studiaţi, şi comportamentul animalelor şi nu au fost semnalate alte probleme de mobilitate. În concluzie, SP este un produs natural care poate fi utilizat în siguranță atât de adulţi cât şi de copii, având numeroase efecte benefice asupra organismului şi nu in ultimul rând datorită toxicităţii sale foarte reduse. **Cuvinte cheie:** spirulina, toxicitate acută, toxicitate cutanată, toxicitate subcronică, toxicitate orală

THE ROLE OF *SPIRULINA PLATENSIS* IN THE CONTROL OF TYPE 2 DIABETES MELLITUS

MARIA-CORINA SERBAN, GHEORGHE STOICHESCU-HOGEA, CAMELIA GURBAN, FLORINA PETCU, DINESHA JEYAKUMAR, FLORINA ANDRICA, CALIN MUNTEANU, SIMONA DRAGAN

University of Medicine and Pharmacy "Victor Babes", Timisoara, Romania

ABSTRACT

In recent years, many experimental and clinical studies evaluated blue-alga *Spirulina platensis* for the treatment of diabetes mellitus (DM). The objective of this study was to evaluate the effects of Spirulina supplements in the control of type 2 diabetes mellitus. The study included patients aged 30-70 years with type 2 DM for at least two years and that have been treated with a stable dose of Metformin for at least 3 months. This randomized, single blinded on parallel groups study included 30 patients with DM type 2, divided in two groups: 15 patients treated with Metformin and placebo (group 1) and 15 patients treated with Metformin and Spirulina (group 2). Spirulina was administered twice per day during 2 months, in a total dose of 800 mg daily. The following parameters were evaluated: blood pressure, total cholesterol (TC), triglycerides (TG), LDL-cholesterol, HDL-cholesterol, fasting serum glucose, and glycated hemoglobin (HbA1c). After 2 months, the mean glycemic value significantly decreased from 124.5 ± 29.28 mg/dl to 108.3 ± 12.53 mg/dl (*p*<0.003) in group 1 and from 149.5 ± 19.89 mg/dl to 121.1 ± 10.66 mg/dl in group 2 (*p*<0.001).The mean value of HbA1c significantly decreased from $7.2\pm0.63\%$ to $6.3 \pm 0.69\%$ in group 1 (*p*=0.008) and from $7.1\pm0.92\%$ to $6.6 \pm 0.33\%$ in group 2 (*p*=0.009).

Keywords: Spirulina platensis, lipids, type 2 diabetes mellitus

INTRODUCTION

Diabetes Mellitus (DM) is a chronic disease that requires long term medical attention to limit and manage the development of devastating complications [1]. An early initiation of pharmacological therapy was associated with improved glycemic control and reduced long term complications in type 2 DM. Many experimental and clinical studies have shown the beneficial effects of food supplements in DM and, particularly, the beneficial effects of Spirulina platensis in the control of type 2 DM. Spirulina is a filamentous cyanobacterium worldwide used as a food supplement [2]. Spirulina platensis (Arthrospira platensis) is one of the three species investigated most often [3]. Spirulina has many nutritional components and it is a rich source of phycocyanin, vitamins, especially vitamin B12 and provitamin A (ß-carotene), minerals, and carotenoids [4]. The lack in cellulose of cell walls in favor of easy digestion is another great advantage of Spirulina consumption [5].

Many experimental and clinical studies have shown the beneficial metabolic, antiviral, liver-protecting, blood-vessel relaxing, anti-cancer, anti-inflammatory and antioxidant properties of Spirulina supplements [6]. In Korea, a double blind, randomized clinical trial evaluated the effects of Spirulina consumption on lipid parameters, immune markers and antioxidant capacity in elderly men. As compared to placebo, Spirulina users had lower lipid parameters, increased serum interleukin-2, but decreased serum interleukin-6, researchers concluding that Spirulina is suitable as a functional food [7]. Another study conducted in Greece showed the beneficial effects of spirulina supplements on lipid profile in 52 dyslipidemic patients. The lipid fractions total cholesterol (TC), LDL-cholesterol (LDL-C) triglycerides (TG), and non-HDL-cholesterol (non-HDL-C) were significantly reduced after 3 months of Spirulina supplementation, the major reduction being observed in TG levels (-16.3%) in women. However this reduction reached 21.3% in women aged > 47 years was 21.3% and 18.6% in women with TG levels >150 mg/dl [8]. These findings are extremely valuable considering the importance of elevated TG levels as a risk factor for cardiovascular disease in women [9]. This study concluded that Spirulina supplementation at a dose of 1g daily has strong hypolipidemic effects especially on TG concentration,

Received 10th of May 2015. Accepted 6th of June 2015. **Address for correspondence**: Lecturer Maria-Corina Serban, MD, PhD, MSc, Department of Functional Sciences, University of Medicine and Pharmacy "Victor Babes" Timisoara, Eftimie Murgu Square No. 2A, RO-300041, Timisoara, Romania; phone: +40752444900, fax : +40256220479; e-mail: dr.corinaserban@yahoo.com

being proposed as a dietary supplement in dyslipidemic patients [8]. Another study done by Parikh *et al* on type 2 DM patients evaluated the hypolipidemic and hypoglycemic effect of Spirulina [10]. The efficacy of Spirulina supplementation (2g/day for 2 months) in both placebo and Spirulina groups was evaluated by determining serum lipid concentrations, blood glucose levels and HbA1c [10]. Spirulina significantly decreased blood glucose, HbA1c TC, TG and apolipoprotein B levels, while HDL-C and apolipoprotein A1 levels marginally increased. This study concluded that Spirulina supplementation has beneficial effects in controlling glycemia and improving the lipid profile of type 2 DM patients [10].

Despite de evidence, the effects of Spirulina supplementation as add-on therapy in diabetic patients have not been conclusive. The objective of this study was to evaluate the effects of Spirulina supplements on lipid profile and glycemic parameters in patients with type 2 DM.

MATERIAL AND METHODS

This interventional study (800 mg Spirulina/daily) was a randomised, single blinded study on parallel groups with 2 months duration. The patients included in the study were aged 30-70 years, diagnosed with type 2 DM for at least two years and under treatment with a stable dose of Metformin for at least 3 months. The exclusion criteria were BMI > 40 kg/m², any acute or chronic inflammatory disease, cancer, kidney disease, moderate or severe hepatic disease and frequent use of antioxidants and anti-inflammatory. The study included 30 patients with type 2 diabetes mellitus (DM), divided as follows: Group 1: 15 patients with DM type 2 + Metformin+ Placebo and Group 2: 15 patients with DM type 2 + Metformin+ Spirulina supplements.

All subjects were instructed to maintain an isocaloric diet and continue with their eating habits throughout the study. The physical examination, chest radiograph, 12-lead electrocardiogram, anthropometric measurements, blood pressure and laboratory tests was carried out at the beginning and after two months of treatment. A venous blood sample was drawn from an antecubital vein in all subjects after an overnight fast to determine total cholesterol (TC), triglycerides (TG) and HDL-cholesterol and uric acid using standard enzymatic method on a COBAS Integra 400 plus analyzer. Bayer diagnostic kit was used to estimate the blood glucose levels. The glycated haemoglobin was determined by the variant haemoglobin A1c using the high performance liquid chromatography technique, using Randox diagnostic kits. Hypertension was diagnosed according to 2013 European Guidelines for the Management of Arterial Hypertension [11]. Diabetes was defined by the current intake of oral hypoglycemic treatment or use of insulin.

For this interventional clinical trial, FAVISAN Company (www.Favisan.ro) provided spirulina and placebo capsules. During this study all subjects were asked to refrain from any source related supplements other than those provided by the study. The compliance of all patients was evaluated weekly by telephone. Statistical analysis was carried out with the programme SPSS-13, which is a software package used for statistical analysis by comparison of data from two

statistical analysis by comparison of data from two samples, paired t test was used and unpaired t-test was used respectively when appropriate (1 and 2 tail tests). The study was conducted according to the guidelines for

Ine study was conducted according to the guidelines for clinical investigation for the treatment and prevention of diabetes CPMP / EWP / 1080 / 00Rev1 delivered on 14 May 2012 by the European Medicines Agency, respecting the Charter of Fundamental Rights of the EU and WMA Declaration of Helsinki (1964) last amended in 2008. This study was conducted in accordance with the Good Clinical Practice of the European Community, CPMP/ICH/135/95 and the ICH Guidelines for Good Clinical Practice: Consolidated Guidance (GCP E-6, April 1996) / and the Declaration of Helsinki and the applicable local laws and regulations.

RESULTS

The study included 30 patients, divided in two groups, similar in terms of age and sex: 15 patients with DM type 2 treated with Metformin and placebo (group 1) and 15 patients with DM type 2 treated with Metformin and the supplement Spirulina twice daily, in a total dose of 800 mg (group 2). No significant differences between anthropometric measurements, mean blood pressure (BP), lipids and glycemic parameters were observed between the two groups at inclusion in the study (Table I).

 Table I. Characteristics of the two groups at inclusion in the study (n=30)

Studied variables	Group	Mean ±Standard deviation (SD)	<i>p</i> value and significance
Age	DM	61.7 ±6.85	0.956 ^{ns}
Aye	CONTROL	61.6±8.90	0.350
Weigh (kg)	DM	103.1±16.09	0.738 ^{ns}
Weigh (kg)	CONTROL	100.7±22.04	0.750
Height (cm)	DM	168.0±7.23	0.686 ^{ns}
neight (chi)	CONTROL	167.0±5.82	0.000
Body mass index	DM	36.6±6.05	0.861 ^{ns}
Douy mass much	CONTROL	36.2±6.93	0.001
Waist	DM	115.3±8.55	
circumference (cm)	CONTROL	113.1±8.29	0.503 ^{ns}
Visceral fat area	DM	195.3±42.97	0.931 ^{ns}
(cm²)	CONTROL	194.0±37.45	0.93113

Systolic blood	DM	141.9±15.57	0.933 ^{ns}
pressure (mmHg)	CONTROL	142.4±15.80	0.355
Diastolic blood	DM	82.0±8.82	0.712 ^{ns}
pressure (mmHg)	CONTROL	83.2±8.68	0.712.12
Glycemia (mg/dl)	DM	149.5±19.89	0.705 ^{ns}
Giycenna (mg/ui)	CONTROL	124.5±29.28	0.705**
TC (mg/dl)	DM	203.3±64.02	0.266 ^{ns}
re (ilig/ul)	CONTROL	211.25±66.43	0.200
TG (mg/dl)	DM	191.2±72.28	0.580 ^{ns}
i G (ilig/ul)	CONTROL	198.7±85.09	0.500
HDL (mg/dl)	DZ	43.9±12.68	0.702 ^{ns}
HDE (IIIg/al)	CONTROL	45.8±16.37	0.702.10
DL (ma/dl)	DM	121.9±51.31	0.259 ^{ns}
LDL (mg/dl)	CONTROL	130.6±46.76	0.20913
Urio opid (mg/dl)	DM	6.2±1.73	0.373 ^{ns}
Uric acid (mg/dl)	CONTROL	6.8±1.84	0.08 ^{ns}

Key: ns – non-significant differences (p>0.05) s – Significant differences (p<0.05)

Table II. The comparison between the parameters studied at inclusion and after two months of Spirulina supplementation in the placebo group (n=15) and Spirulina group (n=15)

The variables		PLACEBO	SP	IRULINA
studied	GROUP		GROUP	
	Mean± Standard deviation	p value	Mean± Standard deviation	р value
Weight (kg)	100.7 ±22.04		103.1 ±16.09	
Weight after 2 months	96.5±14.2	0.008s	100.8±14.57	0.001s
Glycemia (mg/dl)	124.5±29.28		149.5±19.89	<0.001
Glycemia after 2 months	108.3±12.53	<0.003s	121.1±10.66	s
HbA1c %	7.2±0.63		7.1±0.92	
HbA1c after 2 months	6.3±0.69	0.008 ^s	6.6±0.33	0.009 ^s
TC (mg/dl)	211.25±66.43	<0.001s	203.3±64.02	0.004s
TC after 2 months	144.58±38.58	<0.0018	168.5±37.58	0.0043
TG (mg/dl)	198.7±85.09	0.02s	191.2±72.28	0.019 ^{ns}
TG after 2 months	139.9±45.09	0.023	154.5±33.71	0.019"
HDL (mg/dl)	45.8±16.37		43.9±12.68	
HDL after 2 months	48.5±15.36	0.04s	45.5±12.45	0.113 ^{ns}
LDL (mg/dl)	130.6±46.76	0.07ns	121.9±51.31	0.059 ^{ns}
LDL after 2 months	119.4±40.45	0.0713	99.3±18.98	0.009/15
Uric acid	6.8±1.84		6.2±1.73	
Uric acid after 2 months	5.58±0.84	0.008 ^s	5.04±0.86	0.471 ^{ns}

Key: $^{\rm ns}$ – non-significant differences (p>0.05) $^{\rm s}$ – Significant differences (p<0.05)

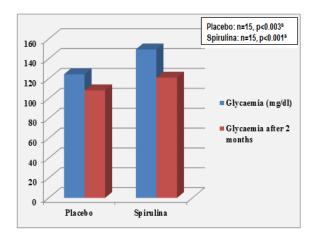
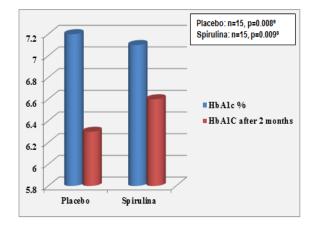
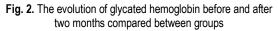


Fig. 1. Evolution of glycemia before and after 2 months compared between groups





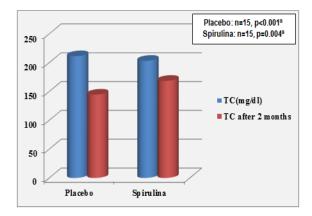


Fig. 3. Evolution of total cholesterol before and after two compared between groups

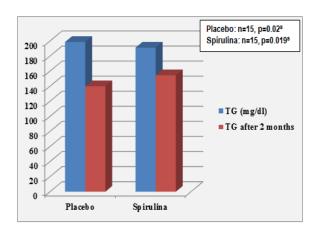


Fig. 4. The evolution of TG before and after two months compared between groups

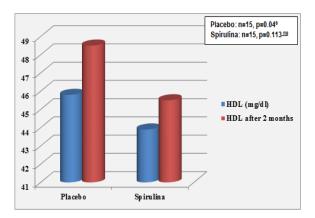


Fig. 5. The evolution of HDL before and after two months compared between groups

DISCUSSION

Our study showed that the supplementation with Spirulina platensis tablets has beneficial effects in controlling blood glucose levels and improving the lipid profile of patients with type 2 DM. The lowering effects of Spirulina on blood lipids are not completely known. Spirulina could act on serum lipids by a similar mechanism as statins, inhibiting the hydroxmethylglutaryl coenzyme A reductase and consecutive reduction of cholesterol synthesis [10]. Although the exact biochemical mechanism by which Spirulina reduces lipid levels is not well understood, some studies have presumed that a high C-phycocyanin content might inhibit the activity of pancreatic lipase activity [7]. The content rich in C-phycocyanin is presumed to act together with glycolipid hemoglobin (Hb)-2 to decrease in jejunal cholesterol absorption and ileal bile acid reabsorption [12]. The results showed that spirulina is probably functioning like an insulin-like protein or may stimulate type beta cells of Langerhans to increase the output of insulin, stimulating the decrease of serum

glucose levels. Spirulina has an effect which leads to the decrease in level of lipoproteins rich in triacylglycerols. In addition, spirulina could act by a similar mechanism as resins, where reabsorption of bile acids in intestine, and cholesterol conversion to bile acids is inhibited [13]. By an unspecified mechanism, spirulina could affect liver cells through which it intervenes directly in cholesterol formation and thus influences its level in serum. Furthermore, spirulina could affect the metabolism of fatty acids, reduce synthesis of VLDL in the liver and as a result minimize the formation of LDL-C in the circulation. Through antioxidative effect, spirulina has shown to reduce lipoperoxidation in liver [14].

Many studies have shown that during diabetes, excess of glucose present in blood reacts with haemoglobin to form HbA1c [15]. The level of HbA1c is always marked as a reliable index of glycaemic control in diabetes. The reason there is an increase in the level of HbA1c is due to decreased level haemoglobin and increased level of blood glucose. The patients who were given spirulina showed a decrease in level of HbA1c, the results in the spirulina group showed an average value of $7.1 \pm 0.92\%$ before use of spirulina and $6.6 \pm 0.33\%$ after, (*p*=0.009), the value of *p* showed a significant difference in HbA1c. The reason for this could be explained by the fact that spirulina is rich in iron and this contributes to the elevated levels of haemoglobin.

Spirulina platensis is considered a safe product, only few patients reported diarrhea after consumption, being Generally Recognized as Safe (GRAS) compound. Our study, in accordance with previous experimental and clinical trials have shown promising data in the control of type 2 DM and therefore it is expected that spirulina, which acts like an insulin like protein can also be used in the future as an anti-diabetic natural substance, eventually replacing oral medication or even insulin.

CONCLUSION

The results obtained in this study showed a significant reduction of weight, glycemia, total cholesterol and triglycerides values after administration of *Spirulina platensis* supplements at dose of 800 mg daily for two months in patients with type 2 DM. These results reveal that *Spirulina platensis* might be a promising dietary supplement for type 2 DM management. Future well-designed larger trials are required to identify the active ingredients beside Phycocyanin and establish the safety profile of *Spirulina platensis* in humans.

COMPETING INTERESTS

The authors declare that they have no competing interests.

ACKNOWLEDGEMENT

This work was supported by the POSDRU grant no. 159/1.5/S/136893: "Strategic partnership for the increase of the scientific research quality in medical universities through the award of doctoral and postdoctoral fellowships – DocMed.Net_2.0".

REFERENCES

1. Allende-Vigo MZ. Diabetes mellitus prevention. *American Journal of Therapeutics*. 2015; 22(1): 68-72.

2. Dmytryk A, Saeid A, Chojnacka K. Biosorption of microelements by Spirulina: towards technology of mineral feed supplements. *The Scientific World Journal.* 2014; 2014: 356328.

3. Holman BW, Malau-Aduli AE. Spirulina as a livestock supplement and animal feed. *Journal of Animal Physiology and Animal Nutrition*. 2013; 97(4): 615-23.

4. Torres-Duran PV, Ferreira-Hermosillo A, Ramos-Jimenez A, Hernandez-Torres RP, Juarez-Oropeza MA. Effect of Spirulina maxima on postprandial lipemia in young runners: a preliminary report. *Journal of Medicinal Food*. 2012; 15(8): 753-7.

5. Dillon J, Phuc A, Dubacq J. Nutritional value of the alga Spirulina. *World Review of Nutrition and Dietetics*. 1994; 77: 32-46.

6. Deng R, Chow TJ. Hypolipidemic, antioxidant, and antiinflammatory activities of microalgae Spirulina. *Cardiovascular Therapeutics*. 2010; 28(4): e33-e45.

7. Park HJ, Lee YJ, Ryu HK, Kim MH, Chung HW, Kim WY. A randomized double-blind, placebo-controlled study to establish

the effects of spirulina in elderly Koreans. Annals of Nutrition & Metabolism. 2008; 52(4): 322-8.

8. Kulshreshtha A, Jarouliya U, Bhadauriya P, Prasad G, Bisen P. Spirulina in health care management. *Current Pharmaceutical Biotechnology.* 2008; 9(5): 400-5.

9. Austin PhD MA, Hokanson M, PhC JE, Edwards PhD KL. Hypertriglyceridemia as a cardiovascular risk factor. *The American Journal of Cardiology*. 1998; 81(4): 7B-12B.

10. Parikh P, Mani U, Iyer U. Role of Spirulina in the control of glycemia and lipidemia in type 2 diabetes mellitus. *Journal of Medicinal Food*. 2001; 4(4): 193-9.

11. Mancia G, Fagard R, Narkiewicz K, Redon J, Zanchetti A, Bohm M, *et al.* 2013 ESH/ESC Guidelines for the management of arterial hypertension: The Task Force for the management of arterial hypertension of the European Society of Hypertension (ESH) and of the European Society of Cardiology (ESC). *European Heart Journal.* 2013; 34(28): 2159-219.

12. Han L-K, Li D-X, Xiang L, Gong X-J, Kondo Y, Suzuki I, et al. Isolation of pancreatic lipase activity-inhibitory component of spirulina platensis and it reduce postprandial triacylglycerolemia. Yakugaku zasshi: Journal of the Pharmaceutical Society of Japan. 2006; 126(1): 43-9.

13. Khan Z, Bhadouria P, Bisen P. Nutritional and therapeutic potential of Spirulina. *Current Pharmaceutical Biotechnology*. 2005; 6(5): 373-9.

14. Ferreira-Hermosillo A, Torres-Duran PV, Juarez-Oropeza MA. Hepatoprotective effects of Spirulina maxima in patients with non-alcoholic fatty liver disease: a case series. *Journal of Medical Case Reports.* 2010; 4(1): 103.

15. Torres-Duran PV, Ferreira-Hermosillo A, Juarez-Oropeza MA. Antihyperlipemic and antihypertensive effects of Spirulina maxima in an open sample of Mexican population: a preliminary report. *Lipids Health Dis.* 2007; 6: 33.

ROLUL SPIRULINA PLATENSIS ÎN CONTROLUL DIABETULUI ZAHARAT TIP 2

REZUMAT

În ultimii ani, numeroase studii experimentale și clinice au evaluat alga albastră *Spirulina platensis* pentru tratamentul diabetului zaharat (DZ). Scopul acestui studiu a fost evaluarea efectelor suplimentelor de Spirulină în controlul DZ de tip 2. Studiul a inclus pacienți cu vârste de 30-70 ani, cu DZ tip 2 diagnosticat de cel puțin doi ani, care au fost tratați cu o doză stabilă de Metformin pentru cel puțin 3 luni. Acest studiu randomizat, singur orb pe grupuri paralele a inclus 30 de pacienți cu DZ tip 2, 15 de pacienți tratați cu Metformin și placebo (grupul 1) și 15 pacienți tratați cu Metformin și Spirulină (grupul 2). Spirulina a fost administrată de două ori pe zi timp de 2 luni, într-o doză totală de 800 mg pe zi. Următorii parametri au fost evaluați: tensiunea arterială, colesterolul total (TC), trigliceridele (TG), LDL-colesterolul, HDL-colesterolul, valorile serice ale glicemiei și hemoglobinei glicozilate (HbA1c). După 2 luni, s-a observat că valoarea medie a glicemiei a scăzut semnificativ de la 124,5 ± 29,28 mg/dl la 108,3 ± 12,53 mg/dl (p <0,003) în grupul 1 și de la 149,5 ± 19,89 mg/dl la 121,1 ± 10,66 mg/dl în grupul 2 (p <0,001). Valoarea medie a HbA1c scăzut semnificativ de la 7,2 ± 0,63% la 6,3 ± 0,69% în grupul 1 (p = 0,008) și de la 7,1 ± 0,92% până la 6,6 ± 0,33 % în grupul 2 (p = 0,009).

Cuvinte cheie: Spirulina platensis, lipide, diabet zaharat tip 2

EFFECTS OF RESVERATROL AND COENZYME Q10 SUPPLEMENTATION IN METABOLIC SYNDROME

FLORINA ARDELEAN^{*}, GHEORGHE STOICHESCU-HOGEA, CAMELIA GURBAN, MARIA-CORINA ŞERBAN, FLORINA PETCU, DIANA S. ANTAL, SIMONA DRĂGAN University of Medicine and Pharmacy "Victor Babes" Timisoara

ABSTRACT

Resveratrol and coenzyme Q10 are two nutraceuticals intensively studied for their benefits in several diseases. Their lipid-lowering, anti-inflammatory and antioxidant activities could improve the condition of patients with metabolic syndrome. Our study aimed to evaluate the effects of the association of a dietary supplement containing resveratrol and coenzyme Q10 to the prescribed medication in patients with metabolic syndrome and statin therapy. The study included 29 patients randomized in two groups, one receiving the dietary supplement (n=15) and one receiving placebo (n=14). The serum levels of total cholesterol (TC), triglycerides (TG), LDL-cholesterol (LDL-C), HDL-cholesterol (HDL-C), transaminases and high sensitivity C-reactive protein (hsCRP) have been evaluated at the beginning of the study and after three months. A decrease of serum levels of TC, TG, LDL-C and hsCRP and an increase of HDL-C were noticed. The greater increase of HDL-C in the group receiving the dietary supplement (from 43.80±13.18 to 46.67±10.40 mg/dl) almost reached a statistical significance (p=0.057). The results indicate that the association of resveratrol and coenzyme Q10 in patients with metabolic syndrome could moderately decrease TC, LDL-C, TG and significantly increase HDL-C. Further studies are needed to elucidate the effects of resveratrol combined with coenzyme Q10 on lipid profile parameters.

Key words: metabolic syndrome, resveratrol, coenzyme Q10

INTRODUCTION

Nowadays, metabolic syndrome (MS) is a condition with an increased prevalence, determining a high risk for cardiovascular disease and type 2 diabetes mellitus (DM). Many factors such as the excess calorie diet, the sedentary life habits, and genetic factors could contribute to the development of this disease [1,2]. Several definitions have been proposed for this complex disease by different organizations. The criteria proposed in these definitions for the diagnosis of MS present similarities, but insulin resistance, lipid and blood pressure (BP) levels are present in most of them [3,4]. The central obesity, dyslipidemia, high BP, proinflammatory and prothrombotic state and insulin resistance are the main factors responsible for the evolution to cardiovascular disease (CVD) and type 2 DM [5]. The chronic low-grade inflammation state in metabolic syndrome might be caused by the generation of leptin and adiponectin and different others adipokines by the adipose tissue [6,7]. The main approach in controlling the risk factors of MS is represented by lifestyle changes (increased physical activity, anti-atherogenic diet) [1]. When lifestyle changes

are insufficient, the drug intervention combines lipid-lowering, hypotensive and hypoglycemic drugs in MS patients [8]. The polymedication in the same patient increases costs, decreases the compliance and is responsible for many side effects [5]. One of the commonly prescribed classes of drugs to control dyslipidemia in MS patients is represented by statins, leading to a reduction of cardiovascular events [9]. Many side effects like muscle and liver toxicity [10], increase in the incidence of type 2 DM, cognitive impairment have been observed in statin users [11]. It has been shown that statins inhibit cholesterol biosynthesis through decreasing the synthesis of mevalonate and consecutive reduction of coenzyme Q10 levels in the organism [12]. Therefore, an association of statins with coenzyme Q10 supplements might be a good strategy in statin users to improve metabolic parameters [13].

Resveratrol (3, 4', 5 trihydroxystilbene) is a polyphenolic compound that has attracted great interest because of its presence in red wine and its multiple benefits [14-16]. The antioxidant [17], cardioprotective [18], anticancer [19], anti-inflammatory [20], lipid-lowering, anti-hyperglycemic [21] or neuroprotective [22] effects of resveratrol have

Received 16th of April 2015. Accepted 12th of May 2015. **Address for correspondence**: Florina Ardelean, Department of Pharmaceutical Botany, Faculty of Pharmacy, "Victor Babes" University of Medicine and Pharmacy Timisoara, Eftimie Murgu Square No. 2A, RO-300041, Timisoara, Romania; phone/fax: +40256 220479; e-mail: florina_ardelean87@yahoo.com

been explored in many experimental and clinical trials. However, the benefits of resveratrol supplementation are still controversial, some recent studies indicating that resveratrol is overestimated [23].

The aim of our study was to evaluate the effect on lipid and inflammation parameters of a dietary supplement containing both resveratrol and coenzyme Q 10 in MS patients on statin therapy.

MATERIAL AND METHOD

This study was randomized, single-blinded with parallel groups and included 29 patients with MS on statin therapy for at least 3 months. The exclusion criteria from this study were represented by any type of cancer or degenerative disease, moderate to severe renal or hepatic disease, acute or chronic inflammatory disease, pregnancy or breastfeeding, abnormal pulmonary X-ray, abnormal ecography, abnormal ECG, deterioration of hematological, hepatic or renal function, hypersensitivity or allergy to any of the compounds of the product administrated in this study, the use of anti-oxidative and anti-inflammatory substances less than two weeks before the start of the study.

The patients enrolled have been distributed in two groups. During three months, the first group (15 patients) received Procor Forte, 1 capsule/day and the second group (14 patients) received placebo. Procor Forte is a dietary supplement containing coenzyme Q10 (60 mg), resveratrol (15 mg), vitamin B6 (3 mg) and magnesium oxide (250 mg) providing 150 g magnesium. For this interventional clinical trial, Sun Wave Pharma Company provided the dietary supplement and the Favisan Company provided the placebo capsules.

The patients have been diagnosed with MS according to the criteria of NCEP-ATP III, presenting 3 or more of the following: waist circumference > 102 cm (men) or > 88 cm (women), triglycerides \geq 150mg/dl, HDL cholesterol < 40mg/dl (men) or < 50mg/dl (women), blood pressure \geq 130/85 mmHg, fasting glucose \geq 110 mg/dl [8]. Patients had increased cardiovascular risk factors (risk SCORE \geq 5). The prescribed medication including statin therapy was respected during the intervention period. Before the beginning of the study patients respected a wash-out period of two weeks in order to eliminate the effects of other dietary supplements or plant extracts consumed before.

At the beginning of the study and after three months the following parameters have been evaluated: anthropometric parameters: weight, height, waist circumference, body mass index (BMI). The serum levels of total cholesterol, triglycerides, LDL, HDL, transaminases, blood glucose, hsCRP and blood pressure were determined in all patients. Bayer diagnostic kit was used to determine the levels of biochemical parameters.

This study was conducted in accordance with the Good Clinical Practice of the European Community, CPMP/ICH/135/95 and the ICH Guidelines for Good Clinical Practice: Consolidated Guidance (GCP E-6, April 1996) / and the Declaration of Helsinki and the applicable local laws and regulations. Written consent was obtained from the patients included in the study. The study was approved by the Ethics Committee of the Institute of Cardiovascular Diseases.

Statistical analysis was done using with the programme SPSS-13, a software package used for statistical analysis by comparison of data from two samples, paired t test was used and unpaired t-test was used respectively when appropriate (1 and 2 tail tests).

RESULTS

The MS patients on statin therapy included in the study presented similar baseline characteristics regarding age, sex, weight, height, BMI. Of the 29 patients 14 were men and 15 women, 6 male and 9 female in the Procor group and 8 male and 6 female in the placebo group. No significant differences have been noted for age (p = 0.289), weight (p = 0.268), height (p = 0.982), BMI (p = 0.222), waist circumference (p = 0.902) as shown in Table I.

	Table I.	The anthropometric parameters	at inclusion
--	----------	-------------------------------	--------------

Parameter	Group	Mean ± standard Deviation	р	
Age (years)	Procor	57.20±9.29	0.200	
	Placebo	61.57±8.90	0.289	
Weight (kg)	Procor	96.56±14.23	0.200	
	Placebo	100.71±22.04	0.268	
Height (cm)	Procor	167.67±8.32	0.000	
	Placebo	167.00±5.82	0.982	
BMI	Procor	33.53±3.72	0 222	
	Placebo	36.22±6.93	0.222	
Waist circumference	Procor	113.40±9.42	0.000	
(cm)	Placebo	113.14±8.39	0.902	

The majority of the patients included in the study received atorvastatin (10 patients in the Procor group and four patients in the placebo group), followed by rosuvastatin (7 patients in the placebo group and 4 in the Procor group). Five patients included in the studied presented side effects to statins (hepatic cytolysis, artralgia and myalgia).

The values of the evaluated parameters in both groups are presented in Table II. For the Procor group (n = 15) significant improvements have been observed for TC with a decrease from 230.40 ± 56.11 mg/dl to 186.13 ± 31.86 mg/dl, LDL-C from 137 ± 38.68 to 114.13 ± 23.39 mg/dl, TG from 167.53 ± 75.99 to 127.07 ± 38.29 mg/dl, ALT from 51.47 ± 50.07 to 18.55±10.33 and hsCRP from 3.73 \pm 2.19 to 2.67 \pm 1.68 mg/dl. No significant improvement was observed for AST levels. The serum values of HDL-C increased from 43.80 \pm 13.18 mg/dl to 46.67 \pm 10.40 mg/dl (p = 0.057) in Procor group (Table II).

Parameter		Procor group Mean±Std.deviation	р	Placebo group Mean±Std.deviation	р
Total cholesterol	Inclusion	230.40±56.11	0.004	233.93±80.89	0.0000
(mg/dl)	3 months	186.13±31.86	0.001	182.14±47.25	0.0002
	Inclusion	43.80±13.18	0.057	45.93±15.97	0.705
HDL-C (mg/dl)	3 months	46.67±10.40	0.057	45.57±13.96	0.785
	Inclusion	137.00±38.68	0.000	144.50±54.40	0.040
LDL-C (mg/dl)	3 months	114.13±23.39	0.003	119.86±38.36	0.018
TC / /	Inclusion	167.53±75.99	0.000	207.21±81.73	0.0007
TG (mg/dl)	3 months	127.07±38.29	0.003	149.43±48.39	0.0002
ALT (11/1)	Inclusion	51.47±50.07	0.022	36.07±19.84	0.000
ALT (U/L)	3 months	18.55±10.33	0.023	22.79±8.47	0.009
ACT (11/1)	Inclusion	36.27±32.30	0.470	33.57±19.63	0.070
AST (U/L)	3 months	24.13±6.92	0.170	24.43±6.15	0.072
	Inclusion	3.73±2.19	0.002	3.43±1.70	0.001
hsCRP (mg/L)	3 months	2.67±1.68	0.002	2.57±1.16	0.001

 Table II. The parameters in the two groups at inclusion and after 3 months

HDL: high density lipoprotein; LDL: low density lipoprotein; TG: triglyceride; ALT: alanine aminotransferase, AST: aspartate aminotransferase, hs CRP: high sensitivity C-reactive protein

In the placebo group (*n*=14), the serum levels of lipid parameters, transaminases and hsCRP decreased after 3 months of supplementation with placebo: TC decreased from 233.93 ± 80.89 to 182.14 ± 47.25 mg/dl, LDL-C from 144.40 ± 54.40 to 119.86 ±38.36 mg/dl and TG from 207.21 ± 81.73 to 149 ± 48,39 mg/dl, ALT from 36.07 ± 19.84 to 22.79 ± 8.47 and hsCRP from 3.43 ± 1.7 to 2.57 ± 1.16 mg/L. A decrease of serum levels of HDL-C, from 45.93±15.97 mg/dl to 45.57±13.96 has been observed.

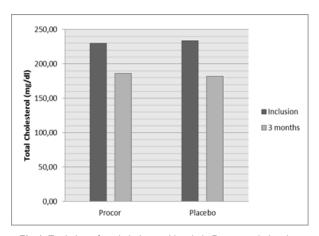


Fig.1. Evolution of total cholesterol levels in Procor and placebo group

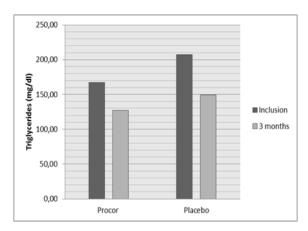


Fig.2. Evolution of TG levels in Procor and placebo group

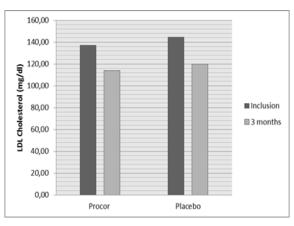


Fig.3. Evolution of LDL-C levels in Procor and placebo group

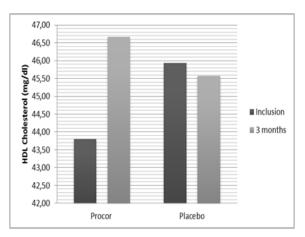
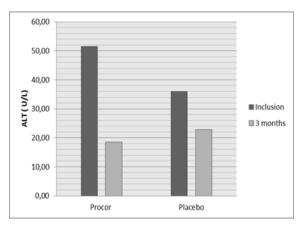


Fig.4. Evolution of HDL-C levels in Procor and placebo group





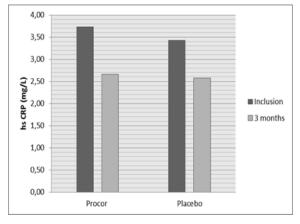


Fig.6. Evolution of hs CRP levels in Procor and placebo group

DISCUSSION

Our study showed positive effects following the administration of a dietary supplement (Procor Forte) containing resveratrol and coenzyme Q10 in patients with MS on statin therapy. The dietary supplement determined an improvement of lipid profile, decreasing TC, LDL-C, and TG and increasing HDL-C (Figures 1-4). The modifications on transaminases and inflammatory parameters like hsCRP were similar in both groups, after Procor or placebo supplementation during three months (Figures 5, 6). The lipid-lowering effects of resveratrol have been noticed in many experimental studies [24,25]. Numerous clinical studies have been performed for resveratrol and coenzyme Q10 to establish the potential benefit of these compounds for health. Even though results indicated by these studies are sometimes controversial, positive effects have been noticed in patients with hypercholesterolemia and type 2 DM [18,26]. A recent study has focused on resveratrol benefits as a dietary supplement in patients with type 2 DM and hypercholesterolemia on statin treatment. The results revealed a decrease of LDL-C, apolipoprotein B

and oxidized LDL after six months of administration with a grape supplement enriched in resveratrol [27]. A decrease of serum levels of LDL-C was also noted after the administration of 10 mg of resveratrol daily in patients with coronary artery disease [18].

Our results are in accordance with previous studies on diabetic patients, which point out an increase of serum levels of HDL-C after supplementation with resveratrol (1g/day) [21]. Moreover, the association of coenzyme Q10 with statin therapy might also increase the serum levels of HDL-C. A study evaluated the benefits of the association of coenzyme Q10 and statins, and revealed increased serum levels of HDL-C in patients on rosuvastatin, compared to patients on atorvastatin [28]. Atorvastatin seems to determine a more evident decrease of ubiquinol levels than rosuvastatin does [28]. Our results indicated an increase of serum levels of HDL-C in the Procor group, supporting the evidence that the association of coenzyme Q10 to statin can improve serum lipid parameters.

The dose of resveratrol present in the dietary supplement was 10 mg. Although other clinical trials have used higher doses of resveratrol, benefits have been observed even al lower doses. Brasnyó et al used in their study capsules containing 5 mg of resveratrol (twice daily) and observed an improvement in insulin sensitivity [29].

The small number of patients included in the study and the short period of intervention could explain the moderate effects of this dietary supplement containing mainly coenzyme Q10 (60 mg) and resveratrol (15 mg) on lipid profile parameters.

CONCLUSION

The association of a dietary supplement containing resveratrol and coenzyme Q10 in patients with metabolic syndrome on statin therapy can improve the lipid profile, mainly through increasing the levels of HDL-C. Further studies conducted on a larger period of time and including an increased number of patients are needed to establish the benefit of the association of resveratrol and coenzyme Q10 in patients wih MS.

COMPETING INTERESTS

The authors declare that they have no competing interests.

ACKNOWLEDGEMENT

This work was supported by the POSDRU grant no. 159/1.5/S/136893: "Strategic partnership for the increase of the scientific research quality in medical universities through the award of doctoral and postdoctoral fellowships – DocMed.Net_2.0" ("Parteneriat strategic pentru creşterea calității cercetării științifice din

universitățile medicale prin acordarea de burse doctorale și postdoctorale – DocMed.Net_2.0").

REFERENCES

1. Kaur J. A Comprehensive Review on Metabolic Syndrome. *Cardiol Res Pract.* 2014; 2014: 943162.

2. Grundy SM. Metabolic syndrome: a multiplex cardiovascular risk factor. *J Clin Endocrinol Metab.* 2007; 92(2): 399-404.

3. Alberti K, Zimmet P, Shaw J. Metabolic syndrome-a new world-wide definition. A consensus statement from the international diabetes federation. *Diabet Med.* 2006; 23(5): 469-80.

4. Dekker JM, Girman C, Rhodes T, Nijpels G, Stehouwer CD, Bouter LM, *et al.* Metabolic syndrome and 10-year cardiovascular disease risk in the Hoorn Study. *Circulation.* 2005; 112(5): 666-73.

5. Grundy SM, Brewer HB, Cleeman JI, Smith SC, Lenfant C. Definition of metabolic syndrome report of the National Heart, Lung, and Blood Institute/American Heart Association Conference on scientific issues related to definition. *Circulation.* 2004; 109(3): 433-8.

6. Kwon H, Pessin JE. Adipokines Mediate Inflammation and Insulin Resistance. *Front Endocrinol (Lausanne).* 2013; 4: 71.

7. Esser N, Legrand-Poels S, Piette J, Scheen AJ, Paquot N. Inflammation as a link between obesity, metabolic syndrome and type 2 diabetes. *Diabetes Res Clin Pract.* 2014; 105(2): 141-50.

8. ATP III P. Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults. National Cholesterol Education Program (NCEP) Adult Treatment Panel III (ATP III). 2001.

9. Jukema JW, Cannon CP, de Craen AJ, Westendorp RG, Trompet S. The controversies of statin therapy: weighing the evidence. *J Am Coll Cardiol.* 2012; 60(10): 875-81.

10. Björnsson E, Jacobsen EI, Kalaitzakis E. Hepatotoxicity associated with statins: reports of idiosyncratic liver injury post-marketing. *J Hepatol.* 2012; 56(2): 374-80.

11. Lewis JH. Clinical perspective: statins and the liver-harmful or helpful? *Dig Dis Sci.* 2012; 57(7): 1754-63.

12. Marcoff L, Thompson PD. The role of coenzyme Q10 in statin-associated myopathy: a systematic review. *J Am Coll Cardiol.* 2007; 49(23): 2231-7.

13. Jiménez-Santos MA, Juárez-Rojop IE, Tovilla-Zárate CA, Espinosa-García MT, Juárez-Oropeza MA, Ramón-Frías T, *et al.* Coenzyme Q10 supplementation improves metabolic parameters, liver function and mitochondrial respiration in rats with high doses of atorvastatin and a cholesterol-rich diet. *Lipids Health Dis.* 2014; 13: 22.

14. Chiva-Blanch G, Arranz S, Lamuela-Raventos RM, Estruch R. Effects of wine, alcohol and polyphenols on cardiovascular disease risk factors: evidences from human studies. *Alcohol Alcohol.* 2013; 48(3): 270-7.

15. Orallo F, Álvarez E, Camiña M, Leiro JM, Gómez E, Fernández P. The possible implication of trans-resveratrol in the cardioprotective effects of long-term moderate wine consumption. *Mol Pharmacol.* 2002; 61(2): 294-302.

16. Vidavalur R, Otani H, Singal PK, Maulik N. Significance of wine and resveratrol in cardiovascular disease: French paradox revisited. *Exp Clin Cardiol.* 2006; 11(3): 217.

17. Carrizzo A, Forte M, Damato A, Trimarco V, Salzano F, Bartolo M, *et al.* Antioxidant effects of resveratrol in cardiovascular, cerebral and metabolic diseases. *Food Chem Toxicol.* 2013; 61: 215-26.

18. Magyar K, Halmosi R, Palfi A, Feher G, Czopf L, Fulop A, *et al.* Cardioprotection by resveratrol: A human clinical trial in patients with stable coronary artery disease. *Clin Hemorheol Microcirc.* 2012; 50(3): 179-87.

19. Aluyen JK, Ton QN, Tran T, Yang AE, Gottlieb HB, Bellanger RA. Resveratrol: potential as anticancer agent. *J Diet Suppl.* 2012; 9(1): 45-56.

20. Udenigwe CC, Ramprasath VR, Aluko RE, Jones PJ. Potential of resveratrol in anticancer and anti-inflammatory therapy. *Nutr Rev.* 2008; 66(8): 445-54.

21. Movahed A, Nabipour I, Lieben Louis X, Thandapilly SJ, Yu L, Kalantarhormozi M, *et al.* Antihyperglycemic effects of short term resveratrol supplementation in type 2 diabetic patients. *Evid Based Complement Alternat Med.* 2013; 2013.

22. Levine C, Li R, Shah Z, Doré S, Watson R, Preedy V. Resveratrol and neurodegenerative diseases. *Botanical medicine in clinical practice*. 2008: 641-50.

23. Hu Y, Liu J, Wang J, Liu Q. The controversial links among calorie restriction, SIRT1, and resveratrol. *Free Radic Biol Med.* 2011; 51(2): 250-6.

24. Rivera L, Morón R, Zarzuelo A, Galisteo M. Long-term resveratrol administration reduces metabolic disturbances and lowers blood pressure in obese Zucker rats. *Biochem Pharmacol.* 2009; 77(6): 1053-63.

25. Robich MP, Osipov RM, Chu LM, Han Y, Feng J, Nezafat R, *et al.* Resveratrol modifies risk factors for coronary artery disease in swine with metabolic syndrome and myocardial ischemia. *Eur J Pharmacol.* 2011;664(1):45-53.

26. Sohet FM, Delzenne NM. Is there a place for coenzyme Q in the management of metabolic disorders associated with obesity? *Nutr Rev.* 2012; 70(11): 631-41.

27. Tomé-Carneiro J, Gonzálvez M, Larrosa M, García-Almagro FJ, Avilés-Plaza F, Parra S, *et al.* Consumption of a grape extract supplement containing resveratrol decreases oxidized LDL and ApoB in patients undergoing primary prevention of cardiovascular disease: A triple-blind, 6-month follow-up, placebo-controlled, randomized trial. *Mol Nutr Food Res.* 2012; 56(5): 810-21.

28. Toyama K, Sugiyama S, Oka H, Iwasaki Y, Sumida H, Tanaka T, et al. Rosuvastatin combined with regular exercise preserves coenzyme Q10 levels associated with a significant increase in high-density lipoprotein cholesterol in patients with coronary artery disease. *Atherosclerosis.* 2011; 217(1): 158-64.

29. Brasnyó P, Molnár GA, Mohás M, Markó L, Laczy B, Cseh J, *et al.* Resveratrol improves insulin sensitivity, reduces oxidative stress and activates the Akt pathway in type 2 diabetic patients. *Br J Nutr.* 2011; 106(03): 383-9.

EFECTELE SUPLIMENTELOR ALIMENTARE CONȚINÂND RESVERATROL ȘI COENZIMA Q10 LA PACIENȚII CU SINDROM METABOLIC

REZUMAT

Resveratrolul și coenzima Q10 sunt două nutraceutice intens studiate pentru beneficiile lor în mai multe boli. Activitățile lor hipolipemiante anti-inflamatorii și antioxidante ar putea îmbunătăți starea pacienților cu sindrom metabolic. Studiul nostru a urmărit evaluarea efectelor asocierii un supliment alimentar care conține resveratrol și coenzima Q10 la medicamentele prescrise la pacienții cu sindrom metabolic și tratament cu statine. Studiul a inclus 29 de pacienți randomizați în două grupuri, unul care a primit suplimentul alimentar (n = 15) și un grup care a primit placebo (n = 14). Nivelurile serice ale colesterolului total (TC), trigliceridelelor (TG), LDL-colesterolului (LDL-C), HDL-colesterolului (HDL-C), transaminazelor și proteinei C reactive cu sensibilitate crescută (hsCRP) au fost evaluate la începutul studiului și după trei luni. S-a observat o scădere a nivelurilor serice ale TC, TG, LDL-C și hsCRP și o creștere a HDL-C. Creșterea HDL-C observată în grupul care a primit suplimentul alimentar (de la 43.80 ± 13.18 la 46.67 ± 10.40 mg / dl), s-a apropiat de valoarea semnificativă din punct de vedere statistic (p = 0,057). Rezultatele prezentului studiu indică faptul că asocierea resveratrolului și coenzimei Q10 la pacienții cu sindrom metabolic ar putea scădea moderat TC, LDL-C, TG și crește semnificativ valoarea HDL-C. Sunt necesare studii suplimentare pentru a elucida efectele combinației resveratrol și coenzima Q10 asupra parametrilor profilului lipidic.

Cuvinte cheie: sindrom metabolic, resveratrol, coenzima Q10

NOVEL INSIGHTS INTO THE ROLE OF METHYLENE BLUE IN MITOCHONDRIAL PROTECTION

OANA M. DUICU^{*}, ADRIAN STURZA^{*}, ILEANA SCURTU-MYTIKO, ANDREEA PRIVISTIRESCU, MARIA DĂNILĂ^{*}, LAVINIA NOVEANU^{*}, DANINA M. MUNTEAN^{*}

Department of Pathophysiology, *Center for Translational Research and Systems Medicine, "Victor Babeş" University of Medicine and Pharmacy, Timişoara, Romania

ABSTRACT

Mitochondrial dysfunction and reactive oxygen species (ROS) generation are critical events in the pathophysiology of ageing and chronic disorders, in the past years modulation of mitochondrial function becoming thus an important therapeutic target. Pharmacological agents able to improve mitochondrial function and/or to decrease ROS production are nowadays highly investigated. Among them an emerging compound is methylene blue (MB), a tricyclic phenothiazine, used for more than a century to treat a variety of disorders. Due to its redox activity, MB has a high affinity for numerous tissue oxidases, including those within mitochondria. Here we briefly review the use of MB in medicine and biology and discuss its beneficial effects as a mild-redox agent able to prevent mitochondria-driven disorders. Methylene blue has a significant potential to improve mitochondrial dysfunction.

Key words: mitochondria, methylene blue, redox potential

INTRODUCTION

Historically, methylene Blue (MB), a tetramethylthione hydrochloride ($C_{16}H_{18}CIN_3S$) is a heterocyclic aromatic compound firstly synthesized by Heinrich Caro and used as a cotton staining dye [1]. In the 19th century, Peter Ehrlich demonstrated its beneficial effects in the treatment of malaria [2]; this report represented the started point for investigating the role of MB in the field of bacterial and viral infections [3] and cancer [3,4]. MB was further used both as therapeutic (e.g., antiseptic, cancer chemotherapy drug) and diagnostic compound (e.g., staining in neuroanatomy and bacteriology, redox agent in biochemical studies) [4,6].

US Food and Drug Administration (FDA) approved MB in treating methemoglobinemia, either inborn [7] or after an overexposure to drugs, nitrophenols, and environmental poisons (e.g., nitrates or cyanides) [8]. The beneficial effects of MB in methemglobinemia therapy are due to its unique autooxidyzing property: indeed, as stipulated by Rojas et al. (2012), "its reduction-oxidation capacity allows electron cycling, without MB gaining any permanent stoichiometric or net reduction" [9](Figure 1). This property explains how MB is acting as an electron donor in the nonenzymatic reduction of methemoglobin: the NADPH methemoglobin reductase reduces MB to LeucoMB

(Figure 1), with a nonenzymatic transfer of electrons to methemoglobin, and by thus restoring the functional hemoglobin and MB [10]. Therefore, same Paul Ehrlich named it the "magic bullet" due to its autooxidyzing property for electron cycling, which was firstly identified in the nervous tissue [11].

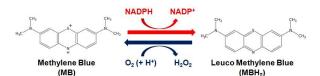


Fig. 1. Methylene blue: a redox active agent. NADPH reduces MB (blue color) to MBH₂ (colorless) which is further oxidized by O₂ (modified after 12).

MB has a high-affinity for the nervous tissue [13], being used firstly in 1938 in the experimental treatment of schizophrenia [14]. Since then, MB has proved to be a neuroprotective agent in numerous neurodegenerative disorders such as Alzheimer's disease (AD), Parkinson's disease (PD), Leber's optic neuropathy [7, 15-22]. The sulfur atom within the structure of MB compound has been suggested to be responsible for this high-affinity to neural structures, yet the underlying mechanism is not completely understood [23].

Received 12th of March 2015. Accepted 17th of April 2015. **Address for correspondence**: Danina Muntean, MD, PhD, Department of Pathophysiology, "Victor Babeş" University of Medicine and Pharmacy of Timişoara, 14 Tudor Vladimirescu st., 300173 Timisoara, RO, Tel/Fax: +40-256-493085; e-mail: daninamuntean@umft.ro

In the past decade, an increasing number of studies investigated the neuroprotective role of MB in various pathological conditions: accordingly, MB reduced the cerebral damage induced by ischemia/reperfusion (I/R) in a model of transient focal cerebral ischemia and attenuated the behavioral and neurochemical impairment in a PD model [24]. Furthermore, MB prevented the decline of the cognitive function in transgenic models of AD and slowed the progression of the disease in a clinical trial [25–27].

MB's protective effects have been also studied in relation with ageing [28] and in pathological states associated with liver failure due to intestinal I/R injury [29]. Thus, MB (at nM levels) was proven to extend the life span of human IMR90 fibroblasts in tissue culture by enhancing mitochondrial function: it increased mitochondrial complex IV, enhanced cellular oxygen consumption, increased heme synthesis, and reversed premature senescence caused by H_2O_2 or cadmium [28]. Collange et al. (2013) demonstrated that the intra-peritoneally injection of MB before reperfusion restored the oxidative capacity of liver mitochondria after I/R injury of the gut, a result which could have beneficial clinical implications during vascular surgery by decreasing the incidence of postoperative multiple organ dysfunction [29].

Nowadays, MB is clinically used in multiple disorders and diagnostic procedures (see Table I).

Table I. Clinical indications of MB and corresponding
dosage (after 25 and 30)

Clinical indications of	MB dosage/Ref.
МВ	
Inherited	1 x 50–250 mg/day (for lifetime) (7)
methemoglobinemia	
Acute	1–2 x 1.3 mg/kg (i.v. over 20
methemoglobinemia	minutes) (30)
Ifosfamid-induced	4 x 50 mg/day p.o. or i.v. (18)
neurotoxicity	
Prevention of urinary tract	3 x 65 mg/day p.o. (30)
infections in elderly	
patients	
Vasoplegic	200 mg i.v. over 1 hour followed by
adrenaline-resistant shock	infusion (0.25–2 mg/kg/hour) (31)
Alzheimer's disease	3 x 60 mg/day (Rember™ according
	to 32)
Pediatric malaria	2 x 12 mg/kg p.o. for 3 days (30)
Parathyroid imaging	3–7.5 mg/kg i.v. (25)
Sentinel lymph node	Local application of 1–5mL of 1% MB
biopsy	(25)

i.v.: intravenous; p.o., oral.

An important approach to improve mitochondrial function is represented by agents with mild redox activity. It is well known that mitochondria are rich in the redox centers (heme, flavin mononucleotide, flavin adenine dinucleotide, iron-sulfur, copper, or natural coenzyme Q) which represent the major source of free radicals production [33]. Thus, since mitochondria physiology depends on major redox activities, redox active agents have been recently proposed as mitochondrial modulators able to enhance energy production and/or to decrease oxidative stress [33].

The mitochondrial electron transport chain (ETC) at the inner mitochondrial membrane is composed of five multimeric complexes (complexes I to V) and two intermediary substrates (coenzyme Q and cytochrome c). The reduced NADH+H⁺ and FADH₂ produced by the intermediate metabolism are further oxidized by the mitochondrial ETC to induce an electrochemical gradient of protons from the mitochondrial matrix to the inter-membrane space (Figure 2), which is finally used by the F₁F₀-ATP synthase (complex V) to produce ATP [34].

Atamna et al. (2012) suggested that redox agents with a mild redox potential (-0.1 to 0.1 V) are able to improve the mitochondrial function via "bypassing the production of superoxide radicals and the cycling between the reduced and oxidized form": mild redox agents enter the cell in the oxidized form, are reduced by the physiologic redox centers (e.g., NADH-dehydronase of complex I), and are further re-oxidized by a second physiologic center (e.g., cytochrome c) [33]. The redox potential of MB is ~ 10 mV, which is considered mild *vs*. the redox potential of the ETC components [28], thereby MB serves as an electron carrier increasing the mitochondrial energy production and inhibiting the superoxide production [35].

As several other drugs, MB presents hormetic pharmacological effects: at low doses elicits beneficial effects, whereas in high concentrations might become deleterious [9]. To enter cytosol, MB is reduced at the cell surface to its lipophilic form, MBH₂. Inside the cell, MBH₂ is re-oxidized to MB by the heme proteins. At low doses, there is an MB-MBH₂ equilibrium (electron cycling) and MBH₂ can donate electrons to ETC complexes and oxygen, leading to enhanced energy metabolism and decreased superoxide formation. At high doses, this MB-MBH₂ equilibrium is impaired and MB can take electrons away from ETC complexes, leading to decreased activity of these complexes and promoting oxidative stress [9].

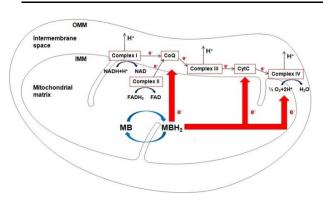


Fig. 2. Schematic representation of methylene blue action within mitochondria (modified after 9). Methylene blue acts as an artificial electron donor to: (i) mitochondria electron transport chain complexes, increasing thus the energy production, and (ii) oxygen, preventing superoxide formation. (IMM - inner mitochondrial membrane; OMM – outer mitochondrial membrane). Illustration realized thanks to Servier Medical Art.

MB hormesis is strongly linked to the mitochondrial membrane potential: a high membrane potential will be responsible for an important accumulation of MB and the dimerization of MB molecules [9]. Gabrielli et al. (2004) demonstrated that oxidative radicals are highly produced in the presence of MB monomers and in a lesser extent in the presence of MB dimers [36]. Thus, low concentrations rather than high concentrations of MB, are associated with enhanced mitochondrial metabolism [37]. In line with these observations, we recently investigated the effects of 5 progressive concentrations of MB (0.05, 0.1, 1, 5, and 10 µM, respectively) on oxygen consumption rate (OCR) and the extracellular acidification rate (ECAR), respectively in H9c2 cardiomyoblasts [38]. We clearly demonstrated the hormetic effect of the acute administration of MB: 0.1 µM induced an increase in both OCR and ECAR, indicating an overall stimulatory impact on cellular bioenergetics. However, all OCR linked parameters and ECAR were decreased in a concentration-dependent manner when MB was administered in the higher concentrations (1, 5, and 10 µM), and were not influenced by 0.05 µM MB, respectively [38]. Our results are in agreement with those reported by Atamna et al. (2008) who showed that human IMR90 fibroblasts in tissue culture exposed to high (micromolar range) but not to low (nanomolar range) concentrations of MB showed a compensatory upregulation of antioxidant enzymes with decreased heme expression and iron uptake [28]. Moreover, high concentrations of MB induced the alteration of neural structure or function in vivo, including humans [37, 39-43]. Another important finding is that not only acute administration of MB is beneficial; chronic administration of low doses of MB increased complex IV activity in brain and heart of old mice, explaining thus the preservation of mitochondrial function and energy metabolism [9, 28, 44, 45].

CONCLUSIONS

It is widely accepted that preventing mitochondrial dysfunction in both acute and chronic pathologies has significant clinical benefits, an observation that rendered mitochondria in the past decades an attractive therapeutic target. In this regard, when administered in an appropriate doses, methylene blue has a significant potential to improve mitochondrial respiration and decrease superoxide radicals formation, via its mild redox potential. Further studies aimed at dissecting the molecular and cellular mechanisms underlying the beneficial effects of methylene blue are definitely warranted.

ACKNOWLEDGMENTS

This work was supported by the POSDRU grant no. 159/1.5/S/136893 titled "Strategic partnership for the increase of the scientific research quality in medical universities through the award of doctoral and postdoctoral fellowships – DocMed.Net_2.0". (O.M.D.). We would like to thank Adrian Wolf (undergraduate student and tutor at the Department of Pathophysiology, "Victor Babes" University of Medicine and Pharmacy, Timişoara) for his valuable contribution to the study.

REFERENCES

1. Caro H. England Patent 3751, 9 October 1877

 Guttmann P, Ehrlich P. U^{*} ber die Wirkung des Methylenblau bei Malaria. *Berlin Klin Woch*. 1891; 28: 953-956.
 Wainwright M. The use of dyes in modern biomedicine. *Biotech Histochem* 2003; 78: 147-155.

4. Wainwright M, Crossley KB. Methylene Blue-a therapeutic dye for all seasons? *J Chemother.* 2002; 14: 431-443.

5. Barbosa P, Peters TM. The effects of vital dyes on living organisms with special reference to methylene blue and neutral red. *Histochem J.* 1971; 3: 71-93.

6. Peter C, Hongwan D, Kupfer A, *et al.* Pharmacokinetics and organ distribution of intravenous and oral methylene blue. *Eur. J. Clin. Pharmacol.* 2000; 56: 247-250.

7. Cawein M, Behlen CH 2nd, Lappat EJ, *et al.* Hereditary diaphorase deficiency and methemoglobinemia. *Arch Intern Med.* 1964; 113: 578-585.

8. Sills MR, Zinkham WH. Methylene blue-induced Heinz body hemolytic anemia. *Arch Pediatr Adolesc Med.* 1994; 148: 306-310.

9. Rojas JC, Bruchey AK, Gonzalez-Lima F. Neurometabolic mechanisms for memory enhancement and neuroprotection of methylene blue. *Prog Neurobiol*. 2012; 96(1): 32-45.

10. Bradberry SM. Occupational methaemoglobinaemia. Mechanisms of production, features, diagnosis and management including the use of methylene blue. *Toxicol Rev.* 2003; 22: 13-27.

11. Ehrlich P. U^{*} ber die Methylenblaureaction der lebenden Nervensubstanz. *Dtsch Med Wochenschr.* 1886; 12: 49-52.

12. Oz M, Lorke DE, Hasan M, *et al.* Cellular and molecular actions of Methylene Blue in the nervous system. *Med Res Rev.* 2011; 31(1): 93-117.

13. Kristiansen JE. Dyes, antipsychotic drugs, and antimicrobial activity. Fragments of a development, with special reference to the influence of Paul Ehrlich. *Dan Med Bull.* 1989; 36: 178-185.

14. Allexsaht WJ. The use of methylene blue in the treatment of catatonic dementia praecox patients. *Psychiatric Quarterly* 1938; 12: 245-252.

15. Naylor GJ, Martin B, Hopwood SE, *et al.* A two-year double-blind crossover trial of the prophylactic effect of methylene blue in manic-depressive psychosis. *Biol Psychiatry.* 1986; 21: 915-920.

16. Kelner MJ, Bagnell R, Hale B, *et al.* Potential of methylene blue to block oxygen radical generation in reperfusion injury. *Basic Life Sci.* 1988; 49: 895-898.

17. Deutsch SI, Rosse RB, Schwartz BL, *et al.* Methylene blue adjuvant therapy of schizophrenia. *Clin Neuropharmacol.* 1997; 20: 357-363.

18. Pelgrims J, De Vos F, Van den Brande J, *et al.* Methylene blue in the treatment and prevention of ifosfamide-induced encephalopathy: report of 12 cases and a review of the literature. *Br J Cancer.* 2000; 82: 291-294.

19. Clifton J 2nd, Leikin JB. Methylene blue. *Am J Ther.* 2003; 10: 289-291.

20. Miclescu A, Basu S, Wiklund L. Methylene blue added to a hypertonic-hyperoncotic solution increases short-term survival in experimental cardiac arrest. *Crit Care Med.* 2006; 34: 2806-13.

21. Miclescu A, Basu S, Wiklund L. Cardio-cerebral and metabolic effects of methylene blue in hypertonic sodium lactate during experimental cardiopulmonary resuscitation. *Resuscitation* 2007; 75: 88-97.

22. Sharma HS, Miclescu A, Wiklund L. Cardiac arrest-induced regional blood-brain barrier breakdown, edema formation and brain pathology: a light and electron microscopic study on a new model for neurodegeneration and neuroprotection in porcine brain. *J Neural Transm.* 2011; 118: 87-114.

23. Müller T: Methylene blue supravital staining. An evaluation of its applicability to the mammalian brain and pineal gland. *Histol Histopathol.* 1998; 13: 1019-26.

24. Wen Y, Li W, Poteet EC, *et al.* Alternative mitochondrial electron transfer as a novel strategy for neuroprotection. *J Biol Chem.* 2011; 286: 16504-15.

25. Oz M, Lorke DE, Petroianu GA. Methylene blue and Alzheimer's disease. *Biochem Pharmacol.* 2009; 78: 927-32.

26. O'Leary JC 3rd, Li Q, Marinec P, *et al.* Phenothiazine-mediated rescue of cognition in tau transgenic mice requires neuroprotection and reduced soluble tau burden. *Mol Neurodegener.* 2010; 5: 45.

27. Medina DX, Caccamo A, Oddo S. Methylene blue reduces abeta levels and rescues early cognitive deficit by increasing proteasome activity. *Brain Pathol.* 2011; 21: 140-149.

28. Atamna H, Nguyen A, Schultz C, *et al.* Methylene blue delays cellular senescence and enhances key mitochondrial biochemical pathways. *Faseb J.* 2008; 22: 703-712.

29. Collange O, Charles AL, Bouitbir J, *et al.* Methylene blue protects liver oxidative capacity after gut ischaemia-reperfusion in the rat. *Eur J Vasc Endovasc Surg.* 2013; 45(2): 168-175.

30. Schirmera RH, Adlera H, Pickhardtb M, *et al.* Lest we forget you - methylene blue. *Neurobiol Aging* 2011; 32(12): 2325.

31. Warth A, Goeppert B, Bopp C, *et al.* Turquoise to dark green organs at autopsy. *Virchows Arch.* 2009; 454: 341-344.

32. Wischik CM, Bentham P, Wischik DJ, *et al.* Tau aggregation inhibitor (TAI) therapy with RemberTM arrests disease progression in mild and moderate Alzheimer's disease over 50 weeks. *Alzheimers Dement.* 2008; 4: T167.

33. Atamna H, Mackey J, Dhahbi JM. Mitochondrial pharmacology: Electron transport chain bypass as strategies to treat mitochondrial dysfunction. *Biofactors* 2012; 38(2): 158-166.

34. Wallace DC. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. *Annu Rev Genet*. 2005; 39: 359-407.

35. Tyler DD. The Mitochondrion in Health and Disease. VHC Publishers, Inc., New York 1992.

36. Gabrielli D, Belisle E, Severino D, *et al.* Binding, aggregation and photochemical properties of methylene blue in mitochondrial suspensions. *Photochem Photobiol.* 2004; 79: 227-232.

37. Vutskits L, Briner A, Klauser P, *et al.* Adverse effects of methylene blue on the central nervous system. *Anesthesiology* 2008; 108: 684-692.

38. Duicu OM, Scurtu I, Popescu R, *et al.* Assessment of the effects of methylene blue on cellular bioenergetics in H9c2 cells. *Rev Chim.* 2015: 66 (4): 519-522.

39. Arieff AJ, Pyzik SW. Quadriplegia after intrathecal injection of methylene blue. *J Am Med Assoc.* 1960; 173: 794-796.

40. Poppers PJ, Mastri AR, Lebeaux M, *et al.* The effect of methylene blue on neural tissue. *Anesthesiology* 1970; 33: 335-340.

41. Blass N, Fung D. Dyed but not dead--methylene blue overdose. *Anesthesiology* 1976; 45: 458-459.

42. Martindale SJ, Stedeford JC. Neurological sequelae following methylene blue injection for parathyroidectomy. *Anaesthesia* 2003; 58:1041-42.

43. Sweet G, Standiford SB. Methylene-blue-associated encephalopathy. *J Am Coll Surg.* 2007; 204: 454-8.

44. Kelner MJ, Bagnell R, Hale B, *et al.* Methylene blue competes with paraquat for reduction by flavo-enzymes resulting in decreased superoxide production in the presence of heme proteins. *Arch Biochem Biophys.* 1988; 262: 422-426.

45. Wrubel KM, Riha PD, Maldonado MA, et al. The brain metabolic enhancer methylene blue improves discrimination learning in rats. *Pharmacol Biochem Behav.* 2007: 86: 712-717.

NOI PERSPECTIVE ASUPRA ROLULUI ALBASTRULUI DE METILEN ÎN PROTECȚIA MITOCONDRIALĂ

REZUMAT

Disfuncția mitocondrială și producerea de specii reactive de oxigen (SRO) constituie evenimente critice în fiziopatologia procesului de îmbătrânire precum și a numeroase afecțiuni cronice, astfel că modularea funcției mitocondriale a devenit o importană țintă terapeutică în ultimii ani. În consecință, agenții farmacologici capabili să îmbunătățească funcția mitocondrială și/sau să inducă reducerea producției de SRO sunt acutalmente intens investigați. Printre aceștia, un compus extrem de important este albastrul de metilen (AM), un compus fenotiazinic triciclic, folosit de mai bine de un secol în tratarea unor afecțiuni diverse. Datorită potențialului său de tip redox, AM prezintă o afinitate crescută pentru numeroase oxidaze tisulare, incuzându-le și pe cele de la nivel mitocondrial. Vom prezenta pe scurt utilizarea AM în medicină și biologie, precum și rolul benefic al AM ca și agent de tip redox capabil de a preveni afecțiunile induse de disfuncția mitocondrială. Albastrul de metilen prezintă un potențial semnificativ de îmbunătățire a metabolismului mitocondrial, reprezentând astfel o abordare terapeutică promițătoare în afecțiunile induse de disfuncția mitocondrială.

Cuvinte cheie: mitocondrii, albastru de metilen, potențial redox